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Genetic diversity of coconut germplasm native to South Kalimantan, Indonesia: a molecular study¹

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ABSTRACT

Coconut (*Cocos nucifera* L.) is one of the most important tree crops in the world, especially in the tropics. This study aimed to determine the genetic diversity of coconut germplasm native to South Kalimantan, Indonesia, using the *rbcL* marker. Nine coconut samples, being eight natively collected from this region and one as an outgroup (obtained from the GenBank database), were used in the study. According to the *rbcL* marker, the coconut germplasm native to South Kalimantan has a relatively high diversity, with a nucleotide diversity (π) of 0.51. The level of diversity is strongly correlated with the mutation present in the observed region, *rbcL*. The phylogenetic analysis showed that the coconut germplasm has a unique relationship, where the 'Dalam' cultivar is the closest to three other dwarf coconuts, i.e., 'Genjah Kuning 1', 'Genjah Kuning 3' and 'Wulung'.

KEYWORDS: *Cocos nucifera* L., breeding program, phylogenetic analysis.

INTRODUCTION

Coconut (*Cocos nucifera* L.), commonly known as the "tree of life" in many nations of the tropics, is one of the most important tree crops in the world (Huang et al. 2013, McKeon et al. 2016). It has currently become a promising export commodity for several countries, e.g., Indonesia and several other Southeast Asian countries (McKeon et al. 2016). In 2020, Indonesia, the Philippines and India were the top three coconut producers worldwide, accounting for 75 % of the global production. Specifically, Indonesia is the largest coconut-producing country, with 17.13 metric tons, or 30 % of the total world production, followed by the Philippines with 14.77 metric tons (26.4 %) and India with 14.68 metric tons (17.0 %) (Burton 2021).

RESUMO

Diversidade genética de germoplasma de coco nativo de Kalimantan do Sul, Indonésia: um estudo molecular

O coco (*Cocos nucifera* L.) é uma das culturas arbóreas mais importantes do mundo, principalmente nos trópicos. Objetivou-se determinar a diversidade genética de germoplasma de coco nativo de Kalimantan do Sul, Indonésia, usando o marcador *rbcL*. Nove amostras de coco, sendo oito coletadas nativamente nessa região e uma como grupo externo (obtida do banco de dados GenBank), foram utilizadas no estudo. Segundo o marcador *rbcL*, o germoplasma de coco nativo de Kalimantan do Sul apresenta diversidade relativamente alta, com diversidade de nucleotídeos (π) de 0,51. O nível de diversidade está fortemente correlacionado com a mutação presente na região observada, *rbcL*. A análise filogenética mostrou que o germoplasma de coco caracteriza-se por uma relação única, onde a cultivar 'Dalam' é a mais próxima a três outros cocos anões, ou seja, 'Genjah Kuning 1', 'Genjah Kuning 3' e 'Wulung'.

PALAVRAS-CHAVE: *Cocos nucifera* L., programa de melhoramento, análise filogenética.

Genetically, the coconut belongs to the *Cocos* monotypic genus, Butiinae subtribe, Cocoeae tribe, Arecoideae subfamily, with unique characteristics (Arunachalam 2012). It is a diploid species ($2n = 32$), with a large genome size of 2.15 Gb nucleotide pairs (Batugal et al. 2009, Arunachalam 2012).

Based on the types of palm trees bearing the fruit, the coconut has two primary varieties, namely 'tall' and 'dwarf' types. The tall varieties, sometimes called var. *typica* (Nar.), are more heterogeneous, due to their cross-pollinated nature (Arunachalam 2012). The dwarf ones, sometimes called var. *nana* (Griff.), are predicted to be mutants from tall types with shorter stature (about 8-10 m, when 20 years old). In nature, they are predominantly self-pollinated, thereby homogenous, and present at the lowest frequency (Arunachalam 2012). According to Huang

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et al. (2013), the dwarf variety, which is less common than the tall one, is usually found growing close to humans and has traits that likely result from human selection.

Nowadays, the major coconut growing countries focus on breeding superior varieties with high yield, high oil content, disease resistance and tolerance to biotic stresses (Batugal et al. 2009). Hence, to support a coconut-breeding program, it is urgent to characterize the genetic diversity of this germplasm. In general, morphological markers are common in determining the genetic diversity of coconut, such as seed germination time, fruit component analysis, floral biology, pollination behavior and foliar traits (Maurice et al. 2015, Zhang et al. 2021). Biochemical parameters, like foliar polyphenols, proteins and isozymes, have also been applied in many studies in coconut diversity analysis (Rajesh et al. 2015). However, these two markers show many drawbacks, e.g., being limited in number, showing modest levels of polymorphism, low heritability, and being influenced by the developmental stages of the plant and varied environmental factors (Rajesh et al. 2014, Rajesh et al. 2015, Le et al. 2020).

Alternatively, molecular markers offer an attractive option, if compared to morphological and biochemical ones, in characterizing the genetic diversity of coconut. Several molecular markers, such as random amplified polymorphic DNA (RAPD) (Rajesh et al. 2014), simple sequence repeat (SSR) (Loiola et al. 2016, Geethanjali et al. 2018, Mahayu & Taryono 2019) and start codon targeted polymorphism (ScoT) (Rajesh et al. 2015), including amplified fragment length polymorphism (AFLP), restriction fragment length polymorphism (RFLP), inverse sequence-tagged repeat (ISTR) and inter simple sequence repeat (ISSR), have been employed to understand the diversity of this germplasm (Arunachalam 2012).

Further, DNA barcoding is a widely used, valuable and effective tool that enables rapid and accurate identification of plant germplasm (Li et al. 2014), including coconut (Le et al. 2020). This technique uses a short DNA sequence from a standard and agreed-upon position in the plastid genome (Li et al. 2014). According to Jeanson et al. (2011), the *rbcL* is one of the selected DNA barcoding markers used for this purpose. While the *rbcL* has certain limitations, like evolving slowly and showing

the lowest divergence, this marker shows certain advantages. For example, it has high universality, is easy to amplify and has discriminatory power at the family and genus levels (Li et al. 2014, Kang et al. 2017). Presently, *rbcL* has been widely used for phylogenetic analysis within families and subclasses of angiosperms and even among different groups of seed plants (Kang et al. 2017).

This study aimed to determine the genetic diversity of coconut germplasm native to South Kalimantan, Indonesia, using the *rbcL* marker.

MATERIAL AND METHODS

Nine coconut samples (Table 1), being eight natively collected from four locations or regencies of South Kalimantan, Indonesia (Figure 1), and one as an outgroup obtained from the GenBank database (NCBI 2021), were used in this study. Most samples were prepared molecularly at the University of Lambung Mangkurat, Indonesia, from May to September 2021.

Sample preparation includes several activities, i.e., DNA extraction, quantification, amplification, electrophoresis, purification and sequencing. The DNA was extracted using young coconut leaf samples, with a commercial DNA extraction kit (Geneaid Biotech Ltd., Taiwan). The DNA quantification was carried out by a NanoVue UV-VIS spectrophotometer (GE Healthcare, UK) and the DNA amplification using the *rbcL* primers, i.e., *rbcL*-F (5'-ATGTCACCACAAACAGAGACTAAAGC-3') and *rbcL*-R (5'-GTAAAATCAAGTCCACCRCG-3') (Gholave et al. 2017). The PCR reaction used a SimpliAMP Thermocycler PCR (Applied Biosystem, USA), with a total volume of 25 µL, consisting of

Table 1. List of native coconut (*Cocos nucifera* L.) germplasm used in this study.

Coconut samples	Type	Origin
'Dalam'	Tall	Barito Kuala, South Kalimantan
'Genjah Kuning 1'	Dwarf	Banjarbaru, South Kalimantan
'Genjah Kuning 2'*	Dwarf	Barito Kuala, South Kalimantan
'Genjah Kuning 3'	Dwarf	Tanah Laut, South Kalimantan
'Genjah Salak 1'	Dwarf	Banjarbaru, South Kalimantan
'Genjah Salak 2'	Dwarf	Banjar, South Kalimantan
'Genjah Salak 3'*	Dwarf	Barito Kuala, South Kalimantan
'Genjah Wulung'	Dwarf	Barito Kuala, South Kalimantan
Voucher 2153**	Dwarf	Puerto Rico

* Introduction; ** outgroup obtained from the GenBank (accession number KX587647).

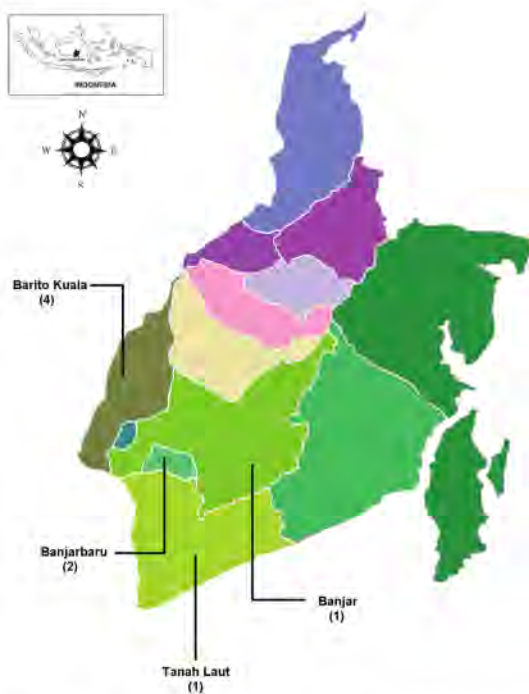


Figure 1. Map of sampling locations (South Kalimantan, Indonesia) where native coconuts were collected. All samples are listed in Table 1.

22 μ L MyTaq HS Red Mix PCR (Bioline, UK), 2.0 μ L (10 μ M) primer forward and reversed, and 1 μ L (10 \times diluted containing ca. ten ng DNA) template. Further, the PCR reaction was programmed with the following conditions: initial denaturation at 94 $^{\circ}$ C for 4 min, denaturation (94 $^{\circ}$ C for 30 sec), annealing (48 $^{\circ}$ C for 30 sec), extension (72 $^{\circ}$ C for 1 min) and final extension (72 $^{\circ}$ C for 7 min) (Mursyidin et al. 2021). Electrophoresis was employed on DNA samples that were successfully amplified using 1.5 % agarose gel, a 1X TBE buffer solution and a FluoroVue stain (SMOBiO Technology, Taiwan). After electrophoresis running, the amplified DNA was observed by a UV transilluminator. DNA purification was carried out on each amplified DNA sample by the criteria for sequencing. DNA sequencing was employed bi-directionally, using the Sanger method by the 1st Base Ltd., Malaysia.

Data analysis began with reconstructing the consensus sequence target, *rbcL*, by the MEGA-X software (Kumar et al. 2018). In this stage, all *rbcL* sequences were checked, refined and assembled

manually. After that, multiple alignments were performed for all *rbcL* sequences by Clustal Omega (Sievers et al. 2020). Third, the phylogenetic analysis of those aligned sequences was performed using the maximum likelihood, neighbor joining and maximum parsimony methods. The internal nodes of all phylogenetic trees were evaluated by the bootstrapping analysis, with 1,000 replicates (Lemey et al. 2009). Finally, the genetic diversity and divergences were determined using Kimura 2-Parameter (K2) distances (Kumar et al. 2018).

RESULTS AND DISCUSSION

Based on the *rbcL* sequence, the coconut germplasm native to South Kalimantan, Indonesia, showed a relatively high diversity, with a nucleotide diversity (π) of 0.51 (Table 2). Several germplasms, e.g., *Amorphophallus* (Gao et al. 2017, Mursyidin & Hemanda 2021, Mursyidin et al. 2022), *Cinnamomum* (Chandrasekara et al. 2021), *Dioscorea* (Ude et al. 2019) and *Elymus* (Dong et al. 2013), have also shown high diversity levels. According to Huang et al. (2016), this level of genetic diversity is often affected by several factors, such as the breeding system, seed dormancy and dispersal mechanism, and geographic variation and range. Life span and other life-history traits, natural selection and the history of populations are other factors that affect the genetic diversity level. Environmental factors are also often responsible for the patterns of its observed diversity at small spatial scales (Huang et al. 2016).

Referred to Lloyd et al. (2016), a high genetic diversity level of the population has a significant impact, both on conservation and breeding programs. Conceptually, it is the main factor in forming a baseline population for natural selection and the

Table 2. Genetic diversity and related information on the *rbcL* sequence of coconut germplasm native to South Kalimantan, Indonesia.

Parameter	Information
Nucleotide diversity (π)	0.51
Number of segregation sites (<i>S</i>)	577
Transition/transversion bias value (<i>R</i>)	0.66
Guanine-Cytosine (GC) content (%)	42.93
Tajima neutrality test (<i>D</i>)	2.42
Bayesian information criterion (BIC)	8,111.01
Akaike information criterion (AICc)	8,006.41
Maximum likelihood value (<i>lnL</i>)	-3,987.15

evolutionary process (Govindaraj et al. 2015). Indeed, it is an essential aspect of the evolutionary trajectory or a precondition for future adaptive changes. Hence, only present-day populations need a high genetic diversity to adapt rapidly (Lloyd et al. 2016).

However, despite the current high genetic diversity, the rapid rate of evolutionary change can exceed this species' adaptation rate. Consequently, future studies are needed to understand how the loss of genetic diversity will affect the ability of future generations to continue to cope with environmental changes in populations (Lloyd et al. 2016). Hence, understanding genetic diversity is very urgent in increasing the effectiveness and efficiency of conservation programs, especially for endangered species. This is because some aspects of conservation biology, such as loss of genetic diversity, are only dealt with in detailed population genetic studies (Luan et al. 2006).

For plant breeding, a level of genetic diversity becomes more urgent in the context of climate change (Govindaraj et al. 2015). Plant breeders use this aspect in developing new and improved cultivars with desirable traits, both associated with various biotic and abiotic stress tolerances (Swarup et al. 2021). It is also essential in generating some agricultural phenomena, like transgressive segregation and heterosis. Diverse hybrid lines are necessary for defect correction of commercial cultivars and the development of new ones. Thus, the identification of cultivars, broadening of diversity and their subsequent utilization are the main goals of future crop improvement or breeding programs (Bhandari et al. 2017).

Further, at a molecular level, a high level of genetic diversity is strongly correlated with the mutation that occurs in the observed region. In this study, polymorphic sites (*S*) had 577 nucleotides (Table 2). According to Frankham et al. (2004), genetic diversity and mutation are two things that are interrelated or cannot be separated from each other. Indeed, the main factor for the emergence of its diversity is mutation (Govindaraj et al. 2015). In other words, the higher the mutation rate, the higher the genetic diversity that it presents. In general, the most common mutations in *rbcL* sequences are substitutions, including transitions and transversions. Insertions/deletions are sometimes present in the region and reveal a conservative nucleotide substitution rate (Clegg 1993).

In this study, the bias value of transition/transversion was 0.66 (Table 2). Referring to Stoltzfus & Norris (2016), this parameter is reflected in the different ratios that affect a complex function of sequence divergence degree. Generally, transitions are more often present in most sequences than transversions (Aloqalaa et al. 2019). Hence, it is common in molecular evolution (Stoltzfus & Norris 2016). This sometimes tends to cause changes in the gene expression or biochemical properties of protein products (Keller et al. 2007).

The Tajima neutrality test (Table 2) indicates that the native coconuts studied were a population that had undergone a balanced selection, and there were no rare alleles because the *rbcL* sequence had $D > 0$. Conceptually, the purpose of the Tajima's test is to identify the region that does not fit the neutral theory model at equilibrium between mutation and genetic drift. In addition, the purpose of the Tajima's *D* test is to distinguish between a DNA sequence evolving randomly ("neutrally") and one evolving under a non-random process, including directional selection or balancing selection, demographic expansion, or contraction, genetic hitchhiking, or introgression. However, to prove this, further studies that are more accurate and comprehensive are necessary (Korneliusson et al. 2013).

The phylogenetic analysis showed that the coconut germplasm native to South Kalimantan has a unique relationship. In general, this germplasm was divided into three clades, based on the maximum likelihood, neighbor joining and maximum parsimony approaches (Figures 2-4). For the maximum likelihood and maximum parsimony (Figures 2 and 4), the 'Dalam' cultivar (which belongs to the tall type of coconut) was grouped into Clade 1, with 'Genjah Kuning 1' and 'Genjah Kuning 3', and 'Genjah Wulung'. Meanwhile, 'Genjah Salak 1' and 'Genjah Salak 3' were grouped into Clade 2. In this case, Clade 3 consisted of two cultivars, i.e., 'Genjah Salak 2' and 'Genjah Kuning 2'. Following the neighbor joining (Figure 3), the sample of voucher 2153 from Puerto Rico (an outgroup) was clustered with (or had the closest relationship to) 'Genjah Kuning 2'.

Further analysis (divergence) showed that the 'Dalam' cultivar had the closest relationship with three other dwarf coconuts, i.e., 'Genjah Kuning 1', 'Genjah Kuning 3' and 'Wulung' (Table 3). Similarly, all three cultivars ('Genjah Kuning 1', 'Genjah Kuning 3' and 'Wulung') are also closely related to

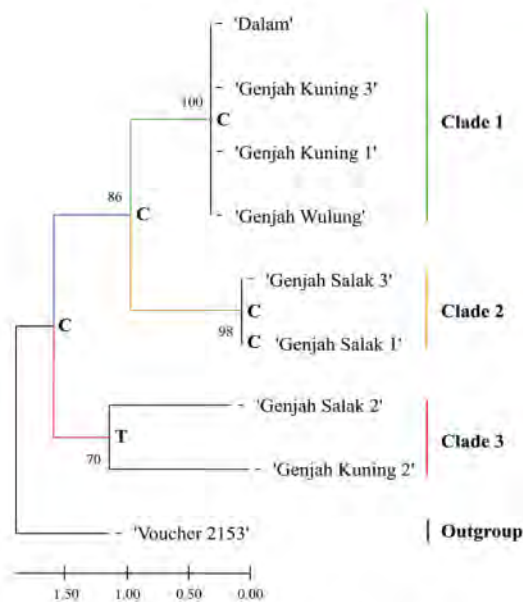


Figure 2. Phylogenetic relationship of coconut germplasm native to South Kalimantan, Indonesia, generated by maximum likelihood and bootstrapping of 1,000 replicates.

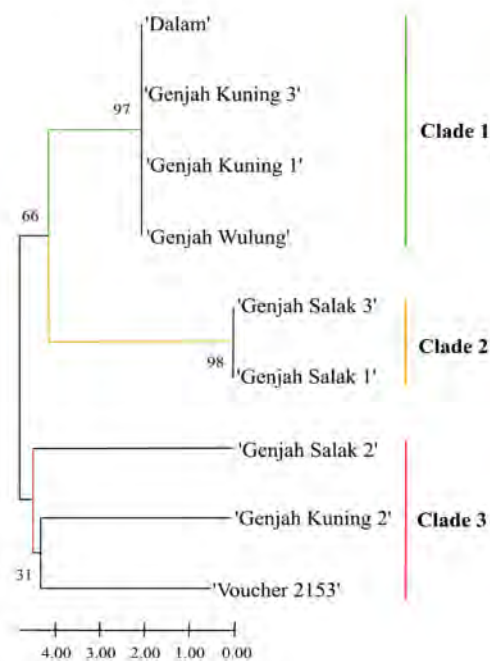


Figure 3. Phylogenetic relationship of coconut germplasm native to South Kalimantan, Indonesia, generated by neighbor joining and bootstrapping of 1,000 replicates.

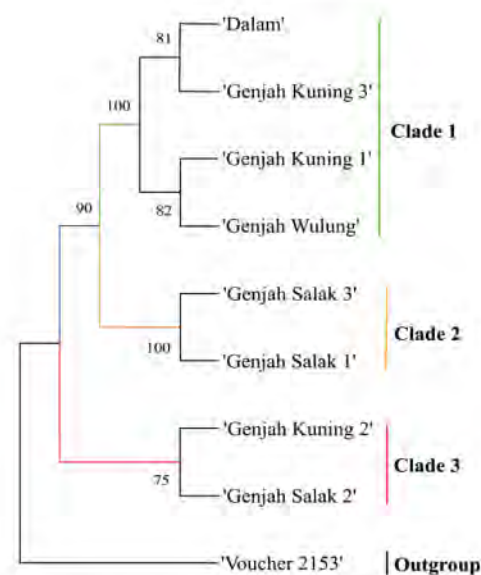


Figure 4. Phylogenetic relationship of coconut germplasm native to South Kalimantan, Indonesia, generated by maximum parsimony and bootstrapping of 1,000 replicates.

one another. Meanwhile, a very distant relationship was shown between 'Genjah Kuning 2' and 'Genjah Salak 3'. Based on morphological characteristics, especially fruit shape and color, 'Genjah Salak' and 'Wulung' are closely related, only differing by the coir color. Besides, based on the same morphological markers, 'Genjah Kuning' is relatively distantly related to the other two cultivars, namely 'Genjah Salak' and 'Wulung'.

Interestingly, following Figure 2, the native coconuts from this region are grouped into paraphyletic and monophyletic groups. In this case, Clades 1 and 2 are monophyletic, whereas Clade 3 is paraphyletic. Initially, all clades have the same common ancestor, with a characteristic C base in the studied region. However, specifically for Clade 3, it changed genetically or molecularly to base T. The bootstrapping analysis confirmed that the grouping of coconut germplasm did not change in general, because the bootstrap value shown on the phylogram was more than 50%, except for the phylogenetic tree generated by neighbor joining, especially between local cultivars with outgroups.

In brief, information on phylogenetic relationships is necessary for researchers, conservationists and

Table 3. Estimates of evolutionary divergence among *rbcL* sequences of coconut native to South Kalimantan, Indonesia.

Operational taxonomic units	Code	A	B	C	D	E	F	G	H	I
'Dalam'	A	0.00								
'Genjah Kuning 1'	B	0.00 ^a	0.00							
'Genjah Kuning 2'	C	3.48	3.48	0.00						
'Genjah Kuning 3'	D	0.00 ^a	0.00 ^a	3.44	0.00					
'Genjah Salak 1'	E	2.98	2.98	4.87	3.03	0.00				
'Genjah Salak 2'	F	3.63	3.63	4.41	3.59	4.44	0.00			
'Genjah Salak 3'	G	2.92	2.92	5.07 ^b	2.97	0.01	4.61	0.00		
'Genjah Wulung'	H	0.00 ^a	0.00 ^a	3.48	0.00 ^a	2.98	3.63	2.92	0.00	
Voucher 2153*	I	3.44	3.44	3.83	3.46	4.12	3.90	3.94	3.44	0.00

* Outgroup; ^a very closely related; ^b farthest related.

breeders (Flint-Garcia 2013). All use the information in predicting the genetic diversity of the offspring when individuals cross or hybridize (Acquaah 2007). In addition, this information may be applied to inferring species and their evolutionary history and helping to analyze the gene flow, genetic differentiation and species delimitation (Fernández-García 2017).

CONCLUSIONS

1. Following the *rbcL* marker, the coconut germplasm native to South Kalimantan, Indonesia, has a relatively high diversity, shown by a nucleotide diversity (π) of 0.51. The level of diversity is strongly correlated with the mutation present in the observed region, *rbcL*;
2. The phylogenetic analysis showed that the coconut germplasm native to this region has a unique relationship. In this case, the 'Dalam' cultivar has the closest relationship with three other dwarf coconuts, i.e., 'Genjah Kuning 1', 'Genjah Kuning 3' and 'Wulung'.

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REFERENCES

ACQUAAH, G. *Principles of plant genetics and breeding*. Oxford: Blackwell, 2007.

ALOQALAA, D. A.; KOWALSKI, D. R.; BLAZEJ, P.; WNETRZAK, M.; MACKIEWICZ, D.; MACKIEWICZ, P. The impact of the transversion/transition ratio on the optimal genetic code graph partition. *In: INTERNATIONAL JOINT CONFERENCE ON BIOMEDICAL ENGINEERING SYSTEMS AND TECHNOLOGIES*, 12., 2019, Prague. *Proceedings...* Setúbal: SciTePress, 2019, p. 55-65.

ARUNACHALAM, V. *Genomics of cultivated palms*. London: Elsevier, 2012.

BATUGAL, P.; BOURDEIX, R.; BAUDOUIN, L. Coconut breeding. *In: JAIN, S. M.; PRIYADARSHAN, P. M. (ed.). Breeding plantation tree crops: tropical species*. New York: Springer Science + Business Media, 2009. p. 327-374.

BHANDARI, H. R.; BHANU, A. N.; SRIVASTAVA, K.; SINGH, M. N.; SHREYA; HEMANTARANJAN, A. Assessment of genetic diversity in crop plants: an overview. *Advances in Plants & Agriculture Research*, v. 7, n. 3, e00255, 2017.

BURTON, J. *The world leaders in coconut production*. 2021. Available at: <https://www.worldatlas.com/articles/the-world-leaders-in-coconut-production.html>. Access on: Nov. 20, 2021.

CHANDRASEKARA, C. H. W. M. R. B.; NARANPANAWA, D. N. U.; BANDUSEKARA, B. S.; PUSHPAKUMARA, D. K. N. G.; WIJESUNDERA, D. S. A.; BANDARANAYAKE, P. C. G. Universal barcoding regions, *rbcL*, *matK* and *trnH-psbA* do not discriminate *Cinnamomum* species in Sri Lanka. *PLoS ONE*, v. 16, n. 2, e0245592, 2021.

CLEGG, M. T. Chloroplast gene sequences and the study of plant evolution. *PNAS*, v. 90, n. 2, p. 363-367, 1993.

DONG, Z. Z.; FAN, X.; SHA, L. N.; ZENG, J.; WANG, Y.; CHEN, Q.; KANG, H. Y.; ZHANG, H. Q.; ZHOU, Y. H. Phylogeny and molecular evolution of the *rbcL* gene

- of *St* genome in *Elymus sensu lato* (Poaceae: Triticeae). *Biochemical Systematics and Ecology*, v. 50, n. 1, p. 322-330, 2013.
- FERNÁNDEZ-GARCÍA, J. L. *Phylogenetics for wildlife conservation*. London: IntechOpen, 2017.
- FLINT-GARCIA, S. A. Genetics and consequences of crop domestication. *Journal of Agricultural and Food Chemistry*, v. 61, n. 35, p. 8267-8276, 2013.
- FRANKHAM, R.; BALLOU, J. D.; BROSCOE, D. A. *A primer of conservation genetics*. Cambridge: Cambridge University Press, 2004.
- GAO, Y.; YIN, S.; YANG, H.; WU, L.; YAN, Y. Genetic diversity and phylogenetic relationships of seven *Amorphophallus* species in southwestern China revealed by chloroplast DNA sequences. *Mitochondrial DNA Part A: DNA Mapping, Sequencing, and Analysis*, v. 29, n. 5, p. 679-686, 2017.
- GEETHANJALI, S.; ANITHA RUKMANI, J.; RAJAKUMAR, D.; KADIRVEL, P.; VISWANATHAN, P. L. Genetic diversity, population structure and association analysis in coconut (*Cocos nucifera* L.) germplasm using SSR markers. *Plant Genetic Resources: Characterization and Utilization*, v. 16, n. 2, p. 156-168, 2018.
- GHOLAVE, A. R.; PAWAR, K. D.; YADAV, S. R.; BAPAT, V. A.; JADHAV, J. P. Reconstruction of molecular phylogeny of closely related *Amorphophallus* species of India using plastid DNA marker and fingerprinting approaches. *Physiology and Molecular Biology of Plants*, v. 23, n. 1, p. 155-167, 2017.
- GOVINDARAJ, M.; VETRIVENTHAN, M.; SRINIVASAN, M. Importance of genetic diversity assessment in crop plants and its recent advances: an overview of its analytical perspectives. *Genetics Research International*, v. 2015, e31487, 2015.
- HUANG, W.; ZHAO, X.; ZHAO, X.; LI, Y.; LIAN, J. Effects of environmental factors on genetic diversity of *Caragana microphylla* in Horqin Sandy Land, Northeast China. *Ecology and Evolution*, v. 6, n. 22, p. 8256-8266, 2016.
- HUANG, Y. Y.; MATZKE, A. J. M.; MATZKE, M. Complete sequence and comparative analysis of the chloroplast genome of coconut palm (*Cocos nucifera*). *PLoS ONE*, v. 8, n. 8, e74736, 2013.
- JEANSON, M. L.; LABAT, J. N.; LITTLE, D. P. DNA barcoding: a new tool for palm taxonomists? *Annals of Botany*, v. 108, n. 8, p. 1445-1451, 2011.
- KANG, Y.; DENG, Z.; ZANG, R.; LONG, W. DNA barcoding analysis and phylogenetic relationships of tree species in tropical cloud forests. *Scientific Reports*, v. 7, n. 1, e12564, 2017.
- KELLER, I.; BENSASSON, D.; NICHOLS, R. A. Transition-transversion bias is not universal: a counter example from grasshopper pseudogenes. *PLoS Genetics*, v. 3, n. 2, p. 185-191, 2007.
- KORNELIUSSEN, T. S.; MOLTKE, I.; ALBRECHTSEN, A.; NIELSEN, R. Calculation of Tajima's D and other neutrality test statistics from low depth next-generation sequencing data. *BMC Bioinformatics*, v. 14, e289, 2013.
- KUMAR, S.; STECHER, G.; LI, M.; KNYAZ, C.; TAMURA, K. MEGA X: molecular evolutionary genetics analysis across computing platforms. *Molecular Biology and Evolution*, v. 35, n. 6, p. 1547-1549, 2018.
- LE, D. T.; ZHANG, Y. Q.; XU, Y.; GUO, L. X.; RUAN, Z. P.; BURGESS, K. S.; GE, X. J. The utility of DNA barcodes to confirm the identification of palm collections in botanical gardens. *PLoS ONE*, v. 15, n. 7, e0235569, 2020.
- LEMEY, P.; SALEMI, M.; VANDAMME, A. M. *The phylogenetic handbook: a practical approach to phylogenetic analysis and hypothesis testing*. Cambridge: Cambridge University Press, 2009.
- LI, X.; YANG, Y.; HENRY, R. J.; ROSSETTO, M.; WANG, Y.; CHEN, S. Plant DNA barcoding: from gene to genome. *Biological Reviews of the Cambridge Philosophical Society*, v. 90, n. 1, p. 157-166, 2014.
- LLOYD, M. M.; MAKUKHOV, A. D.; PESPENI, M. H. Loss of genetic diversity as a consequence of selection in response to high PCO₂. *Evolutionary Applications*, v. 9, n. 9, p. 1124-1132, 2016.
- LOIOLA, C. M.; AZEVEDO, A. O. N.; DINIZ, L. E. C.; ARAGÃO, W. M.; CARLOS, C. D.; SANTOS, P. H. A. D.; RAMOS, H. C. C.; PEREIRA, M. G.; RAMOS, S. R. R. Genetic relationships among tall coconut palm (*Cocos nucifera* L.) accessions of the international coconut genebank for Latin America and the Caribbean (ICG-LAC), evaluated using microsatellite markers (SSRs). *PLoS ONE*, v. 11, n. 3, e0151309, 2016.
- LUAN, S.; CHIANG, T. Y.; GONG, X. High genetic diversity vs. low genetic differentiation in *Nouelia insignis* (Asteraceae), a narrowly distributed and endemic species in China, revealed by ISSR fingerprinting. *Annals of Botany*, v. 98, n. 3, p. 583-589, 2006.
- MAHAYU, W. M.; TARYONO. Coconut (*Cocos nucifera* L.) diversity in Indonesia based on SSR molecular marker. *AIP Conference Proceedings*, v. 2099, e020013, 2019.
- MAURICE, E. O.; MUHAMMED, N.; STEPHEN, M. G.; PASCAL, O. O.; FRANCIS, K. M.; EMMANUEL, M.; JAMES, O. O. *In-situ* morphological characterization of coconut in the coastal lowlands of Kenya. *African Journal of Plant Science*, v. 9, n. 2, p. 65-74, 2015.

- MCKEON, T. A.; HAYES, D. G.; HILDEBRAND, D. F.; WESELAKE, R. J. *Industrial oil crops*. Champaign-Urbana: AOCS, 2016.
- MURSYIDIN, D. H.; HERNANDA, M. A. Phylogenetic positions of three *Amorphophallus* species natively growing in the Meratus Mountains, South Kalimantan, Indonesia. *Biodiversitas*, v. 22, n. 5, p. 2821-2828, 2021.
- MURSYIDIN, D. H.; HERNANDA, M. A.; BADRUZSAUFARI. Genetic diversity of elephant foot yam (*Amorphophallus paeoniifolius*) and two other relatives from the Meratus Mountains of South Kalimantan, Indonesia. *Journal of Tropical Biodiversity and Biotechnology*, v. 7, n. 1, ejtbb66231, 2022.
- MURSYIDIN, D. H.; NAZARI, Y. A.; BADRUZSAUFARI; MASMITRA, M. R. D. DNA barcoding of the tidal swamp rice (*Oryza sativa*) landraces from South Kalimantan, Indonesia. *Biodiversitas*, v. 22, n. 4, p. 1593-1599, 2021.
- NATIONAL CENTER FOR BIOTECHNOLOGY INFORMATION (NCBI). *KX587647.1*. 2021. Available at: <https://www.ncbi.nlm.nih.gov/>. Access on: Sep. 10, 2021.
- RAJESH, M. K.; JERARD, B. A.; PREETHI, P.; THOMAS, R. J.; KARUN, A. Application of RAPD markers in hybrid verification in coconut. *Crop Breeding and Applied Biotechnology*, v. 14, n. 1, p. 36-41, 2014.
- RAJESH, M. K.; SABANA, A. A.; RACHANA, K. E.; RAHMAN, S.; JERARD, B. A.; KARUN, A. Genetic relationship and diversity among coconut (*Cocos nucifera* L.) accessions revealed through SCoT analysis. *3 Biotech*, v. 5, n. 6, p. 999-1006, 2015.
- SIEVERS, F.; BARTON, G.; HIGGINS, D. G. Multiple sequence alignments. In: BAXEVANIS, A. D.; BADER, G. D.; WISHART, D. S. (ed.). *Bioinformatics*. New Jersey: John Wiley & Sons, 2020. p. 227-250.
- STOLTZFUS, A.; NORRIS, R. W. On the causes of evolutionary transition:transversion bias. *Molecular Biology and Evolution*, v. 33, n. 3, p. 595-602, 2016.
- SWARUP, S.; CARGILL, E. J.; CROSBY, K.; FLAGEL, L.; KNISKERN, J.; GLENN, K. C. Genetic diversity is indispensable for plant breeding to improve crops. *Crop Science*, v. 61, n. 2, p. 839-852, 2021.
- UDE, G. N.; IGWE, D. O.; MCCORMICK, J.; OZOKONKWO-ALOR, O.; HARPER, J.; BALLAH, D.; ANINWEZE, C.; CHOSEN, O.; OKORO, M.; ENE, C.; CHIEZE, V.; UNACHUKWU, M.; ONYIA, C.; ACQUAAH, G.; OGBONNA, J.; DAS, A. Genetic diversity and DNA barcoding of yam accessions from southern Nigeria. *American Journal of Plant Sciences*, v. 10, n. 1, p. 179-207, 2019.
- ZHANG, R.; CAO, H.; SUN, C.; MARTIN, J. J. J. Characterization of morphological and fruit quality traits of coconut (*Cocos nucifera* L.) germplasm. *HortScience*, v. 56, n. 8, p. 961-969, 2021.

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