

Genetic diversity and relationship of Durian (*Durio* spp.) Germplasm by the Internal Transcribed Spacer (ITS) Region: In silico analysis

by Dindin Hidayatul Mursyidin

Submission date: 31-May-2023 03:29AM (UTC-0500)

Submission ID: 2105869349

File name: 20-Genetic_Diversity_and_Relationship_of_Durian.pdf (1.04M)

Word count: 5018

Character count: 28263

Genetic Diversity and Relationship of Durian (*Durio* spp.) Germplasm Based on the Internal Transcribed Spacer (ITS) Region: *In Silico* Analysis

[10.18196/planta_tropika.v11i1.13649](https://doi.org/10.18196/planta_tropika.v11i1.13649)

1

Dindin Hidayatul Mursyidin*, Muhammad Irfan Makruf, Muhammad Fitri, Nico Aliannur

4

Laboratory of Genetics and Molecular Biology, Faculty of Mathematics and Natural Sciences, University of Lambung Mangkurat, Jl. A. Yani Km. 36, Banjarbaru, South Kalimantan 70714 Indonesia

*Corresponding author, email: dindinhm@gmail.com

1

Durian (*Durio* spp.) is a germplasm with a relatively high species diversity, with an estimated 27 species worldwide. However, the existence of several species has been threatened. This study aimed to reconstruct the DNA barcode of the durian and its relatives (*Durio* spp.) and analyze the genetic diversity and its relationship based on the internal transcribed spacer (ITS) region. Sixteen sequences of durians ITS were collected from GenBank (NCBI) and analyzed *in silico* using the BLAST, MultAlin, and MEGA-X software, then reconstructed phylogenetically by the UPGMA and Maximum Likelihood methods. The results show that the ITS region of *Durio* spp. has a base length of about 702 bp, where several mutations occur, substitution (transversion and transition) and indel (insertion and deletion). At the nucleotide level, this germplasm shows a relatively high diversity of 0.065. The cluster analyses (UPGMA and Maximum Likelihood) can separate this germplasm into four clusters and five main clades, respectively. In this study, *D. zibethinus*, the most popular species in the *Durio* genus, is closely related to *D. lowianus* and far from *D. griffithii*. This information is beneficial as reference data to support durian conservation and breeding programs, locally and globally, especially in Indonesia.

Keywords: Chloroplast DNA, Breeding, Durian, Genetic diversity, Phylogenetic

ABSTRACT

Durian (*Durio* spp.) merupakan salah satu plasma nutrifah yang memiliki keragaman spesies relatif tinggi, diperkirakan mencapai 27 spesies di seluruh dunia, namun keberadaannya telah terancam. Penelitian ini bertujuan untuk merekonstruksi DNA barcoding durian dan kerabatnya, serta menganalisis keragaman dan kekerabatan genetik secara *in silico* berdasarkan sekuen gen internal transcribed spacer (ITS). Sebanyak 16 sekuen gen ITS *Durio* spp. telah dikoleksi dari GenBank (NCBI) dan dianalisis secara *in silico* menggunakan software BLAST, MultAlin dan MEGA-X, serta direkonstruksi secara filogenetik menggunakan metode UPGMA dan Maximum Likelihood (ML). Hasil penyajian memperlihatkan bahwa region tersebut memiliki panjang basa sekitar 702 bp, yang didalamnya terdapat beberapa peristiwa mutasi, baik substitusi (transversi dan transisi) dan indel (insersi dan delesi). Hasil analisis lebih lanjut menunjukkan bahwa plasma nutrifah ini memiliki keragaman genetik relatif tinggi, sebesar 0.065. Sementara itu, analisis UPGMA dan ML mampu memisahkan plasma nutrifah *Durio* spp., masing-masing kedalam empat kluster dan lima klad utama. Dalam penelitian ini, *D. zibethinus* merupakan spesies paling populer dalam genus *Durio* memiliki kekerabatan sangat dekat dengan *D. lowianus* dan berkerabat jauh dengan *D. griffithii*. Informasi ini diharapkan sangat bermanfaat sebagai data acuan untuk mendukung program pelestarian dan pemuliaan durian (*Durio* spp.), baik secara lokal dan global, terutama di Indonesia.

Kata kunci: DNA kloroplas, Pemuliaan, Durian, Keragaman genetik, Filogenetik

INTRODUCTION

Durian (*Durio* spp.), belonging to the Malvaceae family, is a higher plant with a relatively high diversity of species (Mursyidin et al., 2022). This germplasm is estimated to reach 27 species (Kurniadi et al., 2019). It spreads widely, especially in Indonesia (Sundari et al., 2019). In Indonesia, 20 species of durian are found on several large islands, including Kalimantan (18 species), Sumatra (7 species), Java (1 species), Bali (1 species), Sulawesi (1 species), and Maluku (1 species) (Mursyidin & Daryono, 2016). In Kalimantan, local terms such as *lahong* for *D. zibethinus*, *langsat* for *D. lowianus*, and *langsat* for *D. graveolens* are commonly used. In other regions, such as Thailand, Vietnam, and Malaysia, have named these durians, such as *lahong* for *D. zibethinus*, *langsat* for *D. lowianus*, and *langsat* for *D. graveolens*.



5
Article History
Received: 9 January 2022
Accepted: 28 November 2022



Planta Tropika: Jurnal Agrosains (Journal of Agro Science) is licensed under a Creative Commons Attribution Non-Commercial 4.0 International License.

dulcis, kerantungan for *D. oxleyanus*, and *lai* for *D. kutejensis*. Several others are known as *tupaloh* (*D. acutifolius*), *apun* (*D. excelsus*), *lai kuyu* (*D. griffithii*), and *tekawai* (*D. lowianus*) (Uji, 2004).

In general, most durian species have economically and ecologically essential values (Aziz & Jalil, 2019). For example, nine durian species have edible fruits, namely *D. lowianus*, *D. graveolens*, *D. kutejensis*, *D. oxleyanus*, *D. testudinarium*, *D. grandiflorus*, *D. dulcis*, *Durio excelsus*, and *D. zibethinus* (Aziz & Jalil, 2019). Even *D. zibethinus* is an agricultural commodity with prominent export prospects (Cheon et al., 2017). Indonesia, for example, one of the biggest durian producers in the world, was able to export this fruit to several other countries, including Middle Eastern countries, with a total value of 232,000 USD in 2020 (Rizaty, 2021). In addition, this country produced over 1.19 million metric tons of durian in the same year (Statista Research Department, 2021).

Apart from producing fruit, 14 species of durian also generate wood that can be useful as interior materials. In addition, the bark of several types of durians is also used in medicine, for example, *D. oxleyanus* and *D. griffithii* as malaria drugs because they contain tannin compounds (Feng et al., 2016). However, due to various human activities, such as deforestation and excessive land conversion, especially for plantations, agriculture, settlements, and industry, several durian species have been threatened (Wilcove et al., 2013).

The International Union for Conservation of Nature or IUCN (2021) states that *D. acutifolius*, *D. dulcis*, *D. grandiflorus*, *D. kutejensis*, *D. pinangianus*, and *D. testudinarium* are included as vulnerable, whereas *D. lanceolatus* is the near-threatened. Consequently, employing conservation or preservation, including cultivation and breeding efforts, is indispensable. According to Wintle et al. (2019), conservation is an activity directed at saving and

preserving the existence of endangered species. Meanwhile, breeding/cultivation activities aim to explore and utilize functional genes for developing new superior cultivars (van Huylembroeck, 2018).

In this case, characterization is also urgent to support both activities. However, this activity is performed using morphological markers, which have several limitations because it is time-consuming and highly influenced by environmental factors (Mursyidin & Khairullah, 2020).

The molecular markers provide speed and high accuracy in germplasm characterization activities. Among the existing molecular ones, ITS is a marker with advantages for characterizing germplasm, including durian (Santoso et al., 2017). According to Prahli et al. (2021) and Soumya & Nair (2017), this gene is located between the structural ribosomal RNA (rRNA) of a similar precursor transcript and a non-functional RNA unit with a rapid evolutionary rate. As a result, it can be used to determine germplasm relationships at the genus, species, and subspecies levels (Qin et al., 2017).

Previously, the genetics of durian germplasm have been studied by various molecular markers, such as RAPD (Mursyidin & Daryono, 2016; Prihatini et al., 2016; Hariyati et al., 2013), SSR, and ISSR (Ho et al., 2020; Santoso et al., 2016). However, these markers are highly subjective. In addition, poor consistency, limited repeatability, and complicated operation limit their effectiveness (Wu et al., 2021). This study aimed to reconstruct

a DNA barcoding motif, as well as determine and analyze the genetic diversity and relationship of 16 durian species (*Durio* spp.) *in silico* by utilizing internal transcribed spacer (ITS) gene sequence data provided in GenBank or the National Center for Biotechnology Information (NCBI). According to Sayers et al. (2019), GenBank has a comprehensive database of freely accessible nucleotide sequences or formal gene descriptions. Hence, such a study

does not require high costs or financial support, then can support conservation and breeding activities or germplasm cultivation, particularly durian germplasm (*Durio* spp.), locally and globally (Parikesit et al., 2017).

MATERIALS AND METHODS

This study was conducted from March-May 2020 *in silico* at the Laboratory of Genetics and Molecular Biology, Faculty of Mathematics and Natural Sciences, University of Lambung Mangkurat. This research used internal transcribed spacer (ITS) gene sequence data from 16 durian species (*Durio* spp.) found in GenBank or NCBI (Table 1). In general, this study includes three main activities: nucleotide sequence search and homology (similarity) analysis, nucleotide (multiple) sequence alignment, and analysis of genetic diversity and relationship of those durians obtained.

Nucleotide Sequence Search and Homology Analysis

The internal transcribed spacer (ITS) gene sequence of *D. zibethinus*, which is available on the GenBank or NCBI website (<https://www.ncbi.nlm.nih.gov/>) with accession number MF629779.1, was used as a reference in this study (Table 1). The homology (similarity) analysis with

other durian species, also available on the GenBank website, was then carried out using the BLAST (Basic Local Alignment Search Tool) software. All of the durians (*Durio* spp.) ITS sequences were then copied into text (notepad) format for further analysis.

Multiple Sequence Alignments

Nucleotide (multiple) sequence alignment of the ITS durian region and its relatives (*Durio* spp.) was carried out online using the MultAlin software (Babar et al., 2014). The software is available online at <http://multalin.toulouse.inra.fr/multalin/>.

Analysis of Genetic Diversity and Relationship

Analysis of genetic diversity and the relationship of *Durio* spp. was performed using Molecular Evolutionary Genetics Analysis or MEGA-X software (Kumar et al., 2018). This analysis was started by inputting all ITS durian sequence data (text format) into the MEGA-X software. Before analysis, the sequences were first converted to fasta (.fas) or mega (.meg) format and aligned in the software. After that, genetic diversity analysis was carried out using the nucleotide diversity index (π) method (Nei & Li, 1979). Meanwhile, phylogenetic reconstruction was employed using the UPGMA and Maximum

Table 1. Species of Durians used in this study, the nucleotide length, and GenBank accession number

Species	Nucleotide length (bp)	GenBank Accession Number
<i>D. acutifolius</i>	684	AF287700.1
<i>D. affinis</i>	692	AF287705.1
<i>D. beccarianus</i>	695	AF287707.1
<i>D. carinatus</i>	689	AF287708.1
<i>D. dulcis</i>	689	AF287713.1
<i>D. grandiflorus</i>	683	AF233320.1
<i>D. graveolens</i>	729	MF629770.1
<i>D. griffithii</i>	684	AF233310.1
<i>D. kutejensis</i>	730	MF629750.1
<i>D. lanceolatus</i>	686	AF287709.1
<i>D. lowianus</i>	688	AF287711.1
<i>D. oblongus</i>	692	AF287703.1
<i>D. oxleyanus</i>	688	AF233306.1
<i>D. singaporense</i>	696	AF287701.1
<i>D. testudinarium</i>	694	AF287704.1
<i>D. zibethinus*</i>	747	MF629779.1

*Reference species

Likelihood methods ([Swenson, 2019](#)). The statistical (bootstrap) analysis was then applied to evaluate the internal branches of the phylogenograms ([Kumar et al., 2018](#)).

RESULTS AND DISCUSSION

Multiple Sequence Alignments

Multiple sequence alignment is one of the biological studies which is most frequently used data analysis models ([Shi et al., 2021](#)). This modeling is applied to look at the phylogeny of a whole genome, proteins, identification of horizontally transferred genes, and detection of combined sequences ([Zielezinski et al., 2017](#)). As data sequencing technology advances, the use of this modeling is increasing ([Katoh et al., 2018](#)). The increasing use of multiple sequence alignment modeling has made this area an active research topic, so more than 100 methods have been used. In multiple alignment analysis, all data will be entered from a point in a set of sequences into equivalent classes based on their respective similarities for all members of a common ancestor ([Maiolo et al., 2018](#)).

The durian and its relatives (*Durio* spp.) have a total length of ITS gene sequences of around 702 base pairs (Figure 1), in which several mutation events, both substitutions (transitions and

transversions) and insertion-deletion (indel), are found. Table 2 provides detailed information about mutation events in the ITS durian sequences. There were 217 loci experiencing mutations in the durian ITS gene sequence (Figure 1 and Table 2). Transversion was the most common type of mutation (95 loci) compared to others. Compared to other studies, the number of mutations in durian is higher than in other species. [Soumya & Nair \(2017\)](#) reported that in the ITS region of *Averrhoa* (L.), there were only 87 loci mutations out of a total of 615 bases it had. Similar other cases were shown in *Anoectochilus* ([Thinh et al., 2020](#)), *Aquilaria* ([Lee et al., 2017](#)), *Litsea* ([Fijridiyanto & Murakami, 2019](#)), *Uncaria* ([Zhu et al., 2020](#)), and *Zanthoxylum* ([Zhao et al., 2018](#)).

These study results align with [Soumya & Nair \(2017\)](#), stating that the ITS region is a part of ribosomal RNA with a high evolutionary rate but is universal for different taxa. Moreover, it can be utilized in phylogenetic studies, molecular ecology, detection, and identification of individual pathogens and non-pathogens ([Zhang et al., 2021](#)). According to [Skuza et al. \(2019\)](#), the ITS region has proven to be one of the most informative regions for forming genetic relationships between species in the genus.

Table 2. Mutations on the ITS region of *Durio* spp. germplasm

Mutation type	Number of Mutation
Deletion	13
Insertion	15
Substitution-transition	94
Substitution -transversion	95
Total	217

Table 3. Information of internal transcribed spacer (ITS) of *Durio* spp.*

Parameter	Value
Nucleotide length (bp)	684-747
21† number of the polymorphic site	217
Bayesian information criteria (BIC)	4538.350
Akaike information criteria (AIC)	4320.399
Maximum likelihood v ₂₃ (lnL)	-2130.112
Transition/transversion bias value (R)	1.010
GC content (%)	68.710
Nucleotide diversity (π)	0.065

*Following Kimura 2-parameter model

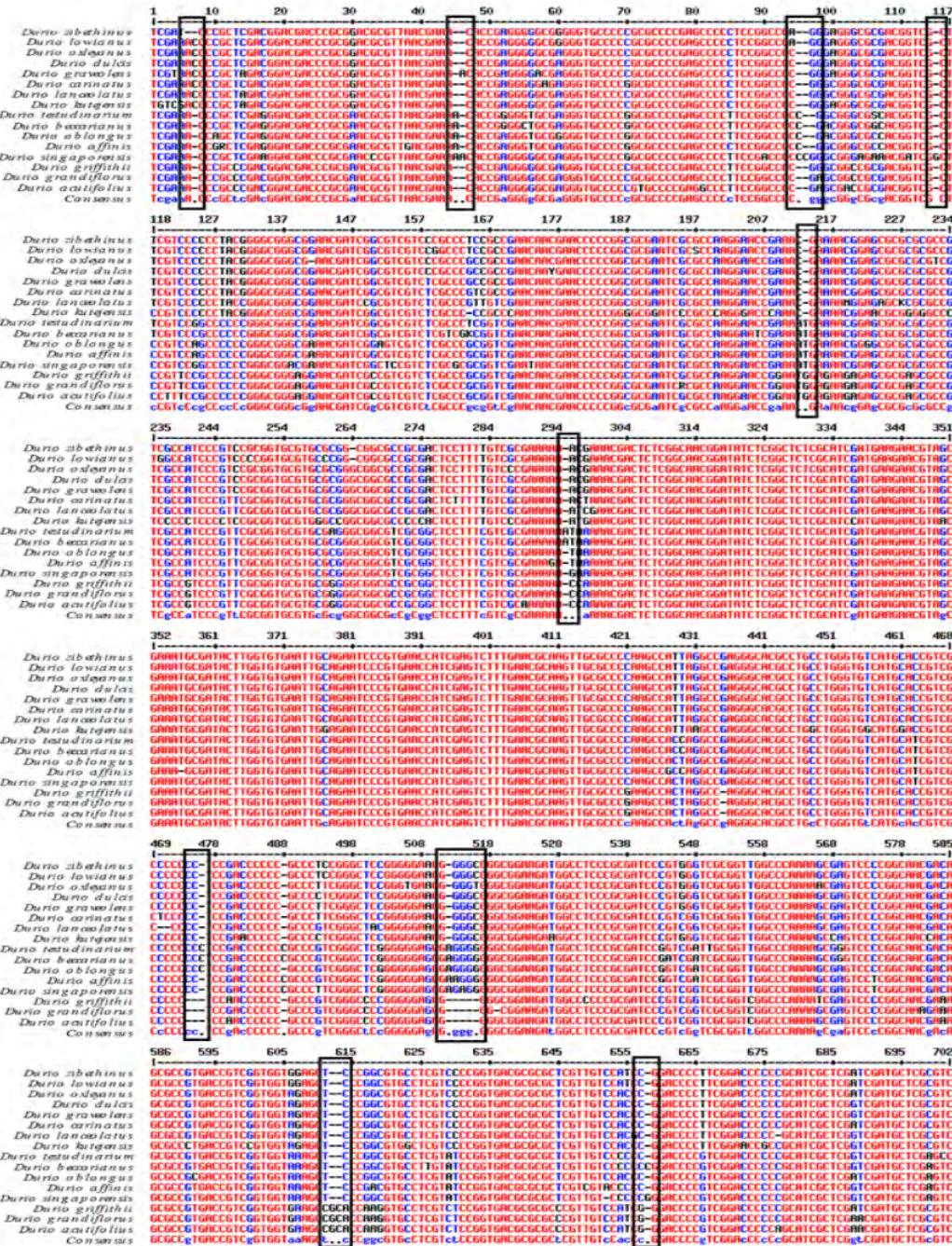


Figure 1. Multiple sequence alignment of the ITS region of *Durio* spp., showing a unique DNA barcoding motif, where mutation, like indels, present therein (close rectangle)

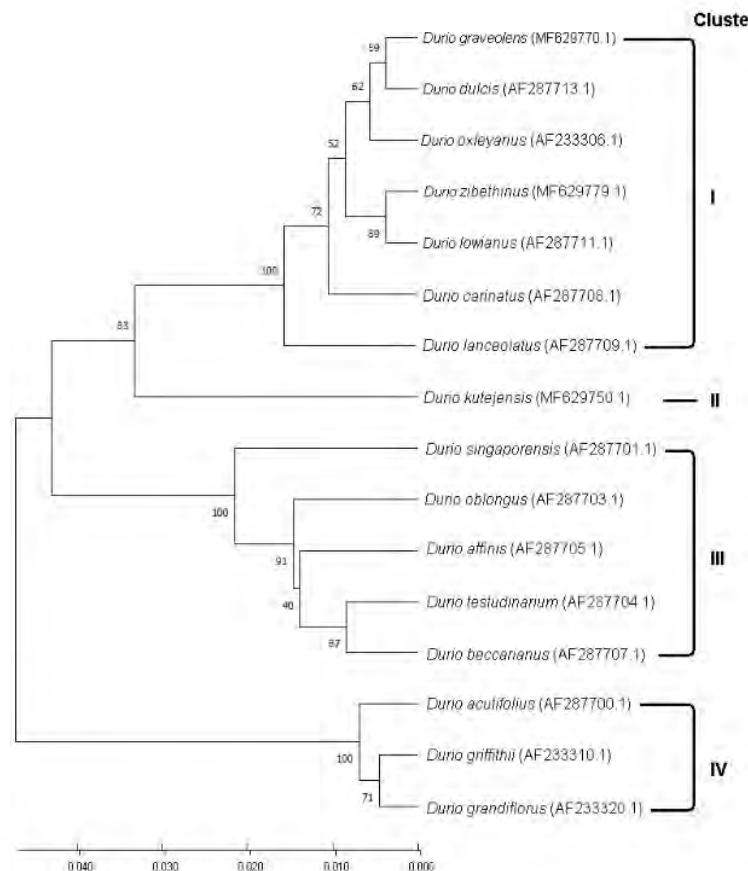


Figure 2. The genetic relationship of *Durio* spp. based on UPGMA analysis. The value on the internal branch shows the results of the bootstrap analysis 1000 replicates

In this case, however, the deletion was the lowest found in the ITS sequence of durians (13 loci). A similar result was found in fungi ([Zhang et al., 2021](#)). According to [Houde et al. \(2019\)](#), such a mutation has a significant value in phylogenetic relations or increases the resolution of evolutionary genetic relationships between the studied taxa. Among candidate DNA barcoding regions, ITS is a non-coding region that generally shows high genetic diversity, including indel polymorphism, so it has the potential ability to be applied in species identification ([Qin et al., 2017](#)).

Genetic Diversity and Relationship of *Durio* spp.

Durian and its relatives (*Durio* spp.) showed relatively high genetic diversity at the nucleotide level, recorded at 0.065 (Table 3). This genetic diversity is closely related to mutation events in the ITS durian sequences studied. Based on Table 3, the *Durio* spp. has ITS sequence character with relatively high polymorphic sites (217 loci), relatively high GC content (68.71%), and the substitution bias value (transition/transversion) is also high (1.01).

According to [van Dorp et al. \(2020\)](#), mutations are the main factor in the emergence of genetic

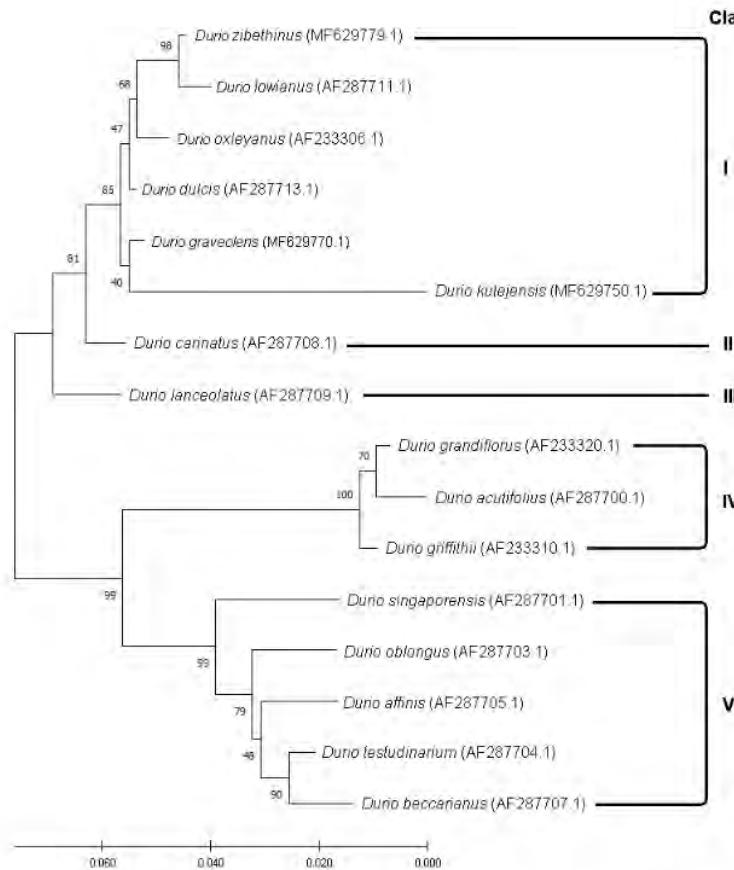


Figure 3. The genetic relationship of *Durio* spp. based on Maximum Likelihood (ML) analysis. The value on the internal branch shows the results of the bootstrap analysis 1000 replicates

9 diversity. In other words, this phenomenon is the primary source that plays a vital role in knowing the differences and evolutionary events between species. In short, a mutation is an event of permanent changes in genetic material, genes, genomes, and chromosomes ([Maluszynski et al., 2017](#)). At the nucleotide (gene or genome) level, mutations include four types, namely deletions, insertions, transitions, and transversions. Deletion and insertion mutations can cause a shift in the coding of the nucleotide triplet. Meanwhile, transition and transversion mutations can change the amino acid

composition formed ([Guo et al., 2017](#)). According to [Qin et al. \(2017\)](#), internal transcribed spacer (ITS) sequences generally have mutations that cause variations between species.

Based on the UPGMA method (Figure 2) and Maximum Likelihood or ML (Figure 3), the durian germplasm (*Durio* spp.) was separated into four clusters and five main clades, respectively. The separation is generally relatively consistent for all species that group together, either into the same cluster or clade. It is strongly related to the bootstrap analysis results and is relatively high in

the resulting internal branches of the dendrogram (Figure 2) and phylogram (Figure 3).

In this study, *D. zibethinus* closely related to *D. lowianus* but very far from *D. griffithii* (see Figures 2 and 3). This study's results align with Nyffeler & Baum (2000) using the ITS marker, stating that *D. zibethinus* is closely related to *D. lowianus*. According to Nyffeler & Baum (2000), these two durian species have morphological similarities, such as calyx, filament arrangement, and anther architecture. Based on other studies, mainly using markers *ndhF* and ITS, a close relationship between *D. zibethinus* and *D. oxleyanus* was reported (Nyffeler & Baum, 2001). Meanwhile, based on the ITS nr-DNA marker, the relationship between *D. zibethinus* and *D. kutejensis* was reported (Santoso et al., 2017). Furthermore, following the UPGMA (Figure 2) and ML (Figure 3), *D. carinatus* and *D. lanceolatus* are nearly related. Naufal (2021) reported that these two durians had the same primitive character in the form of a straight pistil stalk and flowering present on the branches.

However, the relationship between these organisms can show the results of molecular evolution during a time course in the presence of genetic differences (Guerrero et al., 2019). In other words, phylogenetic trees obtained from polymorphisms in a genome, such as chloroplasts, can be used to determine the bar code of an organism and identify differences in taxonomic status and evolution, including population genetics (Nguyen et al., 2017; Xu et al., 2020). Furthermore, information on these relationships benefits the conservation of endangered species and increases plant breeding in general (Bi et al., 2018).

CONCLUSION

Durian germplasm (*Durio* spp.) shows a unique DNA barcode motif based on the internal transcribed spacer (ITS) region sequence. The align-

ment of these sequences shows that the ITS *Durio* spp. has a base length of about 702 bp, in which there are several mutation events, both substitution (transversion and transition) and indel (insertion and deletion). The results also show that this germplasm has a relatively high genetic diversity, amounting to 0.065. Meanwhile, the cluster analysis (UPGMA) and Maximum Likelihood (ML) can separate *Durio* spp. into four clusters and five main clades, respectively. In this study, *D. zibethinus*, the most popular species in the genus *Durio*, is closely related to *D. lowianus* and distantly related to *D. griffithii*. This information is beneficial as reference data to support durian (*Durio* spp.) conservation and breeding programs, both locally and globally, especially in Indonesia.

ACKNOWLEDGMENTS

This study was funded partly by the Director General of Higher Education, Ministry of Education and Culture, Indonesia, through a national student competitive research grant for 2020.

REFERENCES

- Aziz, N. A. A., & Jailil, A. M. M. (2019). Bioactive compounds, nutritional value, and potential health benefits of indigenous durian (*Durio zibethinus* Murr.): A review. *Foods*, 8(3), 1-18. <https://doi.org/10.3390/foods8030096>.
- Babar, M. E., Pervaiz, M. T., Nadeem, A., Hussain, T., & Aslam, N. (2014). Multiple sequence alignment tools: assessing performance of the underlying algorithms. *Journal of Applied Environmental and Biological Sciences*, 4(8S), 76-80.
- Bi, Y., Zhang, M. F., Xue, J., Dong, R., Du, Y. P., & Zhang, H. X. (2018). Chloroplast genomic resources for phylogeny and DNA barcoding: A case study on *Fritillaria*. *Scientific Reports*, 8(1), 1-12. <https://doi.org/10.1038/s41598-018-19591-9>.
- Cheon, S. H., Jo, S., Kim, H. W., Kim, Y. K., Sohn, J. Y. & Kim, K. J. (2017). The complete plastome sequence of Durian, *Durio zibethinus* L. (Malvaceae). *Mitochondrial DNA Part B: Resources*, 2(2), 763-764. <https://doi.org/10.1080/23802359.2017.1398615>.
- Feng, J., Wang, Y., Yi, X., Yang, W. & He, X. (2016). Phenolics from durian exert pronounced NO inhibitory and antioxidant activities. *Journal of Agricultural and Food Chemistry*, 64(21), 4273-4279. <https://doi.org/10.1021/acs.jafc.6b01580>.
- Fijridiyanto, I. A., & Murakami, N. (2019). Evaluating the utility of

- external transcribed spacer (ETS) and internal transcribed spacer sequences (ITS) for phylogenetic analysis of *Litsea* Lam. (Lauraceae) and related genera. *Buletin Kebun Raya*, 22(1), 47-68.
- Guerrero, P. C., Majore, L. C., Cornejo-Romero, A., & Hernández-Hernández, T. (2019). Phylogenetic relationships and evolutionary trends in the cactus family. *Journal of Heredity*, 110(1), 4-21. <https://doi.org/10.1093/hered/esy064>.
- Guo, C., McDowell, I. C., Nodzenski, M., Scholtens, D. M., Allen, A. S., Lowe, W. L., & Reddy, T. E. (2017). Transversions have larger regulatory effects than transitions. *BMC Genomics*, 18(394), 1-9. <https://doi.org/10.1186/s12864-017-3785-4>
- Hariyati, T., Kusnadi, J., & Arumingtyas, E. L. (2013). Genetic diversity of hybrid durian resulted from cross-breeding between *Durio kutejensis* and *Durio zibethinus* based on random amplified polymorphic DNAs (RAPDs). *American Journal of Molecular Biology*, 03, 153-157. <https://doi.org/10.4236/ajmb.2013.33020>.
- Ho, V. T., Ho, M. D., & Tran, T. L. (2020). Characterizing genetic variation of two popular durians (*Durio zibethinus* L.) varieties in southern Vietnam by using ISSR markers. *Bioscience Research*, 17, 3040-3049.
- Houde, P., Braun, E. L., Narula, N., Minjares, U., & Mirarab, S. (2019). Phylogenetic signal of indels and the Neoavian radiation. *Diversity*, 11(7), 1-23. <https://doi.org/10.3390/d11070108>.
- IUCN. (2021). The IUCN Red List of Threatened Species: *Durio*. Retrieved March 27, 2021, from <https://www.iucnredlist.org/>
- Katoh, K., Rozewicki, J., & Yamada, K.D. (2018). MAFFT online service: multiple sequence alignment, interactive sequence choice and visualization. *Briefings in Bioinformatics*, 20(4), 1160-1166. <https://doi.org/10.1093/bib/bbx108>.
- Kumar, S., Stecher, G., Li, M., Knyaz, C., & Tamura, K. (2018). MEGA X: Molecular evolutionary genetics analysis across computing platforms. *Molecular Biology and Evolution*, 35(6), 1547-1549. <https://doi.org/10.1093/molbev/msy096>.
- Kurniadinata, O. F., Wenpei, S., Zaini, A., & Rusdiansyah. (2019). Six potential superior durian plants resulted by cross breeding of *D. zibethinus* and *D. kutejensis* from East Kalimantan, Indonesia: initial identification. *Journal of Tropical Horticulture*, 2(2), 45. <http://dx.doi.org/10.33089/jthort.v2i2.24>.
- Lee, S. Y., Mohamed, R., Faridah-Hanum, I., & Lamasudin, D. U. (2018). Utilization of the internal transcribed spacer (ITS) DNA sequence to trace the geographical sources of *Aquilaria malaccensis* Lam. populations. *Plant Genetic Resources: Characterisation and Utilisation*, 16(2), 103-111. <https://doi.org/10.1017/S1479262117000016>
- Maiolo, M., Zhang, X., Gil, M., & Anisimova, M. (2018). Progressive multiple sequence alignment with indel evolution. *BMC Bioinformatics*, 19(1), 1-8. <https://doi.org/10.1186/s12859-018-2357-1>.
- Maluszynski, M., Szarejko, I., Maluszynska, J., & Szurman-Zubrzcka, M. (2017). Mutation techniques. In *Encyclopedia of Applied Plant Sciences* (Vol. 2, pp. 215-228). Elsevier Inc. <https://dx.doi.org/10.1016/B978-0-12-394802-6.00121-0>.
- Mursyidin, D. H. & Daryono, B. S. (2016). Genetic diversity of local durian (*Durio zibethinus* Murr.) cultivars of South Kalimantan's province based on RAPD markers. *AIP Conference Proceedings*, 1755(040008), 1-7. <https://doi.org/10.1063/1.4958483>.
- Mursyidin, D. H., & Khairullah, I. (2020). Genetic evaluation of tidal swamp rice from South Kalimantan, Indonesia based on the agro-morphological markers. *Biodiversitas Journal of Biological Diversity*, 21(10), 4795-4803. <https://doi.org/10.13057/biodiv/d211045>.
- Mursyidin, D. H., Makruf, M. I., Badruzaufari, & Noor, A. (2022). Molecular diversity of exotic durian (*Durio* spp.) germplasm: a case study of Kalimantan, Indonesia. *Journal of Genetic Engineering and Biotechnology*, 20(39), 1-13. <https://doi.org/10.1186/s43141-022-00321-8>
- Naufal, D.I. (2021). Studi filogenetika *Durio* di Kalimantan berdasarkan karakter morfologi bunga (Unpublished Thesis). Universitas Islam Negeri Syarif Hidayatullah.
- Nei, M., & Li, W.H. (1979). Mathematical model for studying genetic variation in terms of restriction endonucleases. *Proceedings of the National Academy of Sciences*, 76(10), 5269-5273. <https://doi.org/10.1073/pnas.76.10.5269>.
- Nguyen, V. B., Park, H. S., Lee, S. C., Lee, J., Park, J. Y., & Yang, T. J. (2017). Authentication markers for five major *Panax* species developed via comparative analysis of complete chloroplast genome sequences. *Journal of Agricultural and Food Chemistry*, 65(30), 6298-6306. <https://doi.org/10.1021/acs.jafc.7b00925>.
- Nyffeler, R., & Baum, D.A. (2000). Phylogenetic relationships of the durians (Bombacaceae-Durioneae or /Malvaceae/Helicteroidae/Durioneae) based on chloroplast and nuclear ribosomal DNA sequences. *Plant Systematics and Evolution*, 224(1-2), 55-82. <https://doi.org/10.1007/BF00985266>.
- Nyffeler, R., & Baum, D. A. (2001). Systematics and character evolution in *Durio* s. lat. (malvaceae/helicteroidae/durioneae or bombacaceae-durioneae). *Organisms Diversity and Evolution*, 1(3), 165-178. <https://doi.org/10.1028/1439-6092-00015>.
- Parikesit, A. A., Anurogo, D., & Putranto, R. A. (2017). Pemanfaatan bioinformatika dalam bidang pertanian dan kesehatan. *Menara Perkebunan*, 85(2), 105-115. http://dx.doi.org/10.22302/irbb_jurmp.v85i2.237.
- Prahl, R. E., Khan, S., & Deo, R. C. (2021). The role of internal transcribed spacer 2 secondary structures in classifying mycoparasitic Ampelomyces. *PLoS ONE*, 16, 1-28. <https://doi.org/10.1371/journal.pone.0253772>.
- Prihatini, R., Ihsan, F., & Indriyani, N. L. P. (2016). Genomic profiling of F1 hybrids of durian (*Durio zibethinus*) revealed by RAPD-PCR. *Journal of Horticulture Research*, 24, 69-76. <https://doi.org/10.1515/jhr-2016-0022>.
- Qin, Y., Li, M., Cao, Y., Gao, Y., & Zhang, W. (2017). Molecular thresholds of ITS2 and their implications for molecular evolution and species identification in seed plants. *Scientific Reports*, 7(1), 1-8. <https://doi.org/10.1038/s41598-017-17695-2>.
- Rizaty, M.A. (2021). National production of durian [Produksi durian nasional]. Retrieved September 27, 2021, from <https://databoks.katadata.co.id/datapublish/2021/06/23/produksi-durian-di-indonesia-menurun-pada-2020>.
- Santoso, P. J., Granitia, A., Indriyani, N. L. P., & Pancoro, A. (2016). Loci analysis and diversity of durian (*Durio* sp.) germplasm based on microsatellite markers. *Jurnal Hortikultur*, 26, 9-20.
- Santoso, P. J., Indriyani, N. L. P., Istianto, M., Pancoro, A., & Aryanta, I. N. P. (2017). Phylogeny of Indonesian durian (*Durio* sp.) germplasm based on polymorphism of ITS-nrDNA sequences.

- Acta Horticulturae*, 1186, 35-41.
- Sayers, E. W., Mark, C., Karen, C., James, O., Kim, D. P., & Ilenie, K. M. (2019). GenBank. *Nucleic Acids Research*, 47, D94-D99.
- Shi, H., Shi, H., & Xu, S. (2021). Efficient multiple sequences alignment algorithm generation via components assembly under PAR framework. *Frontiers in Genetics*, 11, 1-7. <https://doi.org/10.3389/fgene.2020.628175>.
- Skuza, L., Szułko, I., Filip, E., & Strzala, T. (2019). Genetic diversity and relationship between cultivated, weedy and wild rye species as revealed by chloroplast and mitochondrial DNA non-coding regions analysis. *PLoS ONE*, 14(2), 1-21. <https://doi.org/10.1371/journal.pone.0213023>.
- Soumya, S. L. & Nair, B. R. (2017). Internal transcribed spacer (ITS) sequence analysis of nuclear ribosomal DNA (nrDNA) in *Averrhoa L.* *International Journal of Current Research*, 9(1), 45353-45359.
- 3** Statista Research Department (2021). Production of durian in **3** Indonesia 2011-2020. Retrieved November 05, 2021, from <https://www.statista.com/statistics/706504/production-of-durian-in-indonesia/>
- 8** Sunda **8** Mas'ud, A., Arumintyas, E. L., Hakim, L., Azrianingsih, R., & Wahyudi, D. (2019). Taxonomic status of local durian (*Durio* spp.) from Ternate Island North Maluku based on morphological character and geographical factor. *International Journal of Conservation Science*, 10(4), 711-720.
- Swenson, N. G. (2019). *Phylogenetic ecology: A history, critics & remodelling*. The University of Chicago Press.
- Thinh, B. B., Chac, L. D., & Thu, L. T. M. (2020). Application of internal transcribed spacer (ITS) sequences for identifying *Anoectochilus setaceus* Blume in Thanh Hoa, Vietnam. *Proceedings on Applied Botany, Genetics and Breeding*, 181(2), 108-116. <https://doi.org/10.30901/2227-8834-2020-2-108-116>
- Uji, T. (2004). Keanekaragaman jenis, plasma nutfah, dan potensi buah-buahan asli Kalimantan. *BioSmart*, 6(2), 117-125.
- van Dorp, L., Acman, M., Richard, D., Shaw, L. P., Ford, C. E., Ormond, L., Owen, C. J., Pang, J., Tan, C. C. S., Boshier, F. A. T., Ortiz, A. T., & Balloux, F. (2020). Emergence of genomic diversity and recurrent mutations in SARS-CoV-2. *Infection, Genetics, and Evolution*, 83, 1-9. <https://doi.org/10.1016/j.meegid.2020.104351>.
- van Huylenbroeck J. (2018). *Handbook of Plant Breeding: Ornamental crops*. Springer International Publishing AG.
- Wilcove, D. S., Giam, X., Edwards, D. P., Fisher, B., & Koh, L. P. (2013). Navjot's nightmare revisited: logging, agriculture, and biodiversity in Southeast Asia. *Trends in Ecology and Evolution*, 28(9), 531-540. <https://doi.org/10.1016/j.tree.2013.04.005>.
- Wintle, B. A., Kujala, H., Whitehead, A., Cameron, A., Veloz, S., Kukkala, A., Moilanen, A., Gordon, A., Lentini, P. E., Cadenhead, N. C. R., & Bekessy, S. A. (2019). Global synthesis of conservation studies reveals the importance of small habitat patches for biodiversity. *PNAS*, 116(3), 909-914. <https://doi.org/10.1073/pnas.1813051115>.
- Wu, F., Ma, S., Zhou, J., Han, C., Hu, R., Yang, X., Nie, G., & Zhang, X. (2021). Genetic diversity and population structure analysis in a large collection of white clover (*Trifolium repens* L.) germplasm worldwide. *PeerJ*, 9, 1-17. <https://doi.org/10.7717/peerj.11325>.
- Xu, J., Shen, X., Liao, B., Xu, J., & Hou, D. (2020). Comparing and phylogenetic analysis chloroplast genome of three *Achyranthes* species. *Scientific Reports*, 10(1), 1-13. <https://doi.org/10.1038/s41598-020-67679-y>.
- Zhang, Y. Z., Han, Q. D., Fu, L. W., Wang, Y. X., Sui, Z. H., & Liu, Y. G. (2021). Molecular identification and phylogenetic analysis of fungal pathogens isolated from diseased fish in Xinjiang, China. *Journal of Fish Biology*, 99(6), 1887-1898. <https://doi.org/10.1111/jfb.14893>.
- Zhao, L. L., Feng, S. J., Tian, J. Y., Wei, A. Z., & Yang, T. X. (2018). Internal transcribed spacer 2 (ITS2) barcodes: A useful tool for identifying Chinese *Zanthoxylum*. *Applications in Plant Sciences*, 6(6), e1157. <https://doi.org/10.1002/aps.3.1157>
- 10** Zhu **10** Li, Q., Chen, S., Wang, Y., Zhou, L., Zeng, C., & Dong, J. (2018). Phylogenetic analysis of *Uncaria* species based on internal **18** transcribed spacer (ITS) region and ITS2 secondary structure. *Pharmaceutical Biology*, 56(1), 548-558. <https://doi.org/10.1080/13880209.2018.1499780>
- Zelezinski, A., Vinga, S., Almeida, J., & Karlowski, W. M. (2017). Alignment-free sequence comparison: benefits, applications, and tools. *Genome Biology*, 18(1), 1-17. <https://doi.org/10.1186/s13059-017-1319-7>.

Genetic diversity and relationship of Durian (*Durio* spp.) Germplasm by the Internal Transcribed Spacer (ITS) Region: In silico analysis

ORIGINALITY REPORT



PRIMARY SOURCES

1	doaj.org Internet Source	7%
2	Submitted to Universitas Negeri Surabaya The State University of Surabaya Student Paper	1%
3	jgeb.springeropen.com Internet Source	1%
4	journal.unnes.ac.id Internet Source	1%
5	repository.umpr.ac.id Internet Source	1%
6	psasir.upm.edu.my Internet Source	<1%
7	journal.peradaban.ac.id Internet Source	<1%
8	repository.uin-malang.ac.id Internet Source	<1%

9

repo-dosen.ulm.ac.id

Internet Source

<1 %

10

Marika Kaden, Katrin Sophie Bohnsack, Mirko Weber, Mateusz Kudła, Kaja Gutowska, Jacek Blazewicz, Thomas Villmann. "Learning vector quantization as an interpretable classifier for the detection of SARS-CoV-2 types based on their RNA sequences", *Neural Computing and Applications*, 2021

Publication

<1 %

11

Dindin Hidayatul Mursyidin. "Phylogenetic relationship of superior durian (*Durio zibethinus*) cultivars native to South Kalimantan, Indonesia", *Pesquisa Agropecuária Tropical*, 2022

Publication

<1 %

12

www.researchgate.net

Internet Source

<1 %

13

Jiantang Xu, Aiqing Li, Xiaofei Wang, Jianmin Qi, Liwu Zhang, Guangqing Zhang, Jianguang Su, Aifen Tao. "Genetic diversity and phylogenetic relationship of kenaf (*Hibiscus cannabinus L.*) accessions evaluated by SRAP and ISSR", *Biochemical Systematics and Ecology*, 2013

Publication

<1 %

14

pdfs.semanticscholar.org

Internet Source

<1 %

15 terengganumyheritage.blogspot.com <1 %
Internet Source

16 Dindin Hidayatul Mursyidin, Fajar Nurrahman <1 %
Maulana. "KERAGAMAN DAN KEKERABATAN
GENETIK GARCINIA BERDASARKAN
KANDUNGAN SENYAWA BIOAKTIF DAN
AKTIVITAS BIOLOGISNYA: KAJIAN IN SILICO",
BERITA BIOLOGI, 2020

Publication

17 ddd.uab.cat <1 %
Internet Source

18 repositorio.una.ac.cr <1 %
Internet Source

19 ejournal.forda-mof.org <1 %
Internet Source

20 polarresearch.net <1 %
Internet Source

21 www.collectionscanada.ca <1 %
Internet Source

22 repo.unand.ac.id <1 %
Internet Source

23 www.cropj.com <1 %
Internet Source

24

www.mdpi.com

Internet Source

<1 %

25

www.medrxiv.org

Internet Source

<1 %

26

Dindin Hidayatul Mursyidin, Muhammad Irfan Makruf, Badruzsaufari, Aidi Noor. "Molecular diversity of exotic durian (*Durio* spp.) germplasm: a case study of Kalimantan, Indonesia", Journal of Genetic Engineering and Biotechnology, 2022

Publication

<1 %

Exclude quotes

On

Exclude matches

Off

Exclude bibliography

On