Granulicatella Adiacens Bacteria Isolation from Perodontitical Patients with Polymerase Chain Reaction Techniques

by drg bayuindra

Submission date: 31-Jul-2020 05:54PM (UTC-0400)

Submission ID: 1357369251

File name: Jurnal Granulicatella Adiacens Bacteria Isolation.doc (664.5K)

Word count: 2785

Character count: 20760

Granulicatella Adiacens Bacteria Isolation from Perodontitical Patients with Polymerase Chain Reaction Techniques

Harun Achmad¹, Sri Oktawati², Andi Mardiana Adam², Burhanuddin Pasiga⁵, Rizalinda Sjahril⁴, Aulia Azizah⁵, Bayu Indra Sukmana⁶, Huldani⁻, Heri Siswanto⁶, Ingrid Neormansyah⁶

¹Lecturer of Pedodontics Department, Dentistry Faculty, Hasanuddin University, Makassar, Indonesia

²Lecturer of Periodontology Department, Dentistry Faculty, Hasanuddin University, Makassar, Indonesia

³Lecturer of Dental Public Health Department, Dentistry Faculty, Hasanuddin University, Makassar, Indonesia

⁴Lecturer of Microbiology Department, Medical Faculty, Hasanuddin University, Makassar, Indonesia

⁵Department of Preventive and Public Health Dentistry, Faculty of Dentistry, Lambung Mangkurat University, Banjarmasin, Indonesia.

⁶Department of Dental Radiology, Faculty of Dentistry, Lambung Mangkurat University, Banjarmasin, Indonesia.

⁷Department of Physiology, Faculty of Medicine, Lambung Mangkurat University, Banjarmasin Indonesia.

⁸Resident of Periodontology Department, Dentistry Faculty, Hasanuddin University, Makassar, Indonesia Correspondence Author E-mail: harunachmader@gmail.com

Article History: Submitted: 10.01.2020 Revised: 12.03.2020 Accepted: 24.04.2020

ABSTRACT

Background: Periodontitis is an inflammatory disease of dental support tissue caused by certain groups of microorganisms, resulting in progressive destruction of the periodontal ligament and alveolar bone, with pocket formation, recession, or both¹. Destruction of periodontal caused by the host defence.

Objective: Granulicatella adiacens is a small Gram-positive, non-motile, non-spore-forming, facultatively anaerobic coccus. As a member of Nutritionally Variant Streptococci, genus Granulicatella, it is a part of the normal microbiota in the oral cavity, urogenital, and intestinal tract. It may also participate in periodontitis, caries, and endodontic infections. Our aim in this study was to investigate the presence of Granulicatella adiacens in periodontal disease

Methods: A total 18 samples from patients with chronic periodontitis was collected, by using the conventional PCR technique for checking the presence of bacterial bands, followed by using DNA sequence methods, to complete the examination of bacteria with Conventional PCR which results are used as a basis for nucleotide examination on

the NCBI Gene Bank using BLAST analysis.

Result: In this study, Granulicatella adiancense was found in 13 samples from 18 solated samples from patients with chronic periodontitis and in periodontal pocket depth more than 7 mm.

Conclusion: Granulicatella adiancense is a normal flora bacteria in the oral cavity and chronic periodontitis conditions, the presence of this bacteria is detected in the deeper periodontal pocket depth.

Keywords: Chronic periodontitis, DNA squence, Granulicatella adiacens

Correspondence:

Harun Achmad

Lecturer of Pedodontics Department, Dentistry Faculty, Hasanuddin

Makassar, Indonesia

E-mail: harunachmader@gmail.com

DOI: 10.31838/srp.2020.4.59

@Advanced Scientific Research. All rights reserved

INTRODUCTION

Periodontitis is an inflammatory disease ofdental support tissue caused by certain groups of microorganisms, resulting in progressive destruction of the periodontal ligament and alveolar bone, with pocket formation, recession, or both1. Destruction of periodontal caused by the host defence. Clinically periodontitis is characterized by plaque accumulation, calculus and pocket formation, periodontal tissue inflammation and alveolar bone loss, and gum bleeding. Pathogenic bacteria are suspected of causing an inflammatory response, gingival and periodontal damage2. Periodontal disease can be treated primarily with debridement supragingiva and subgingiva. The combination of surgical or mechanical treatments and proper oral hygiene can maintain a healthy periodontal condition3. Periodontal disease is no longer solely caused by the presence of specific bacteria but by ecological changes in the biofilm community and the population level of bacterial species in the oral microbiota4. Bacteria in the phylum firmicutes are lactic acid bacteria, which is commensal bacteria in the oral cavity. However, due to a change from symbiosis to dysbiosis, this bacteria becomes a pathogen. Lactic acid bacteria have the ability to ferment sugar into lactic acid which can further cause demineralization and bone resorption.5Nutritionally variant streptococcus (NVS) were originally described by Frenkel and Hirsch (1961) as a new type of streptococcus exhibiting satellitism around

colonies of other bacteria. Isolates have been recovered from blood, abscesses, or al ulcers, and urethral samples. Because of both difficulties in culturing these organisms and the variety of appearances that they present on primary detection, such strains have caused major diagnostic difficulties. ⁶

The condition of normal flora is where microorganisms that are usually in certain locations and under normal circumstances do not cause disease⁶. Granulicatella adiacens in the form of small Gram-positive, non-motile, non-spore coccus, oxidase, and catalase-negative, facultative anaerobes. The nutritional requirements of Granulicatella adiacensare complex and require pyridoxal or L-cysteine in growth media as essential nutrition for the normal growth of this bacteria7. Granulicatella adiacenshas a normal commensal relationship on most mucosal surfaces which allows it to affect the mucosal area of the human body but although these bacteria can infect all these areas, these bacteria rarely cause disease species of this genus found in the oral cavity, intestine and urinary tract8. Because of complex nutritional growth requirements, Granulicatella adiacenscan most often cause infections in nutrient-rich tissue, such as the endocardium or the eye, causing endocarditis or endophthalmitis, in addition to causing intravascular infections. Infection in this area causes endovascular infections, the central nervous system, ocular, jawbones and joints, and joint and urinary tract infections9.

Granulicatella infection is quite alarming for several reasons. First, this infection, especially endocarditis, has been associated with greater morbidity and mortality compared with other infections caused by another streptococcus. Second, Granulicatella infection has antibiotic resistance and the recurrence rate is higher than other streptococcus¹⁰. The present study was carried out to determine the presence of Granulicaella adiacens bacteria in chronic periodontitis patients from the subgingival plaque of these patients.

MATERIAL AND METHODS

This study was approved by the ethical committee of the Faculty of Dentistry, Hasanuddin University, Makassar, Indonesia. The study included a total of 18 subjects (8 Male and 10 female) undergoing periodontal treatment. Medical histories from the participants were obtained and thorough clinical and radiographic examinations of their teeth were conducted after written informed consent. Patients with a pocket depth of at least 4 mm; aged 35-50 years systemic disease; OHI-S Moderate-severe; patients do not have a systemic disease; were inclusion criteria and the exclusion criteria were bacteria does not obtain from the sample and bacterial nucleotides do not exist in Gene Bank.

Assessment of periodontal status was performed by a single trained examiner, the status was means of clinical attachment loss (CAL), probing pocket depths (PPD), which was measured more than four millimeters per tooth by using a WHO periodontal probe. Oral hygiene indices such as papillary bleeding index (PBI), plaque index (PI), and gingival index (GI) were also assessed.

Sample Collection: The sample was collected from the gingival area of buccal tooth surfaces using sterile curettes

onto the swab, then the swab put into sterile transport media.

Microbiological Analysis: The swab is inserted into a tube containing the BHIB (Brain Heart Infusion Broth) medium then incubated for at least 8 hours at 370 C. The bacteria culture transferred to Blood Agar medium and incubated for 24 hours at 370 C.

DNA preparation: The basic principle of DNA extraction is a series of processes to separate DNA from other components such as proteins and others. This extraction procedure includes the process of Sample Preparation, Cell Lysis, DNA Binding, Wash, Elution. The liquid contained in the Eppendorf tube is a DNA product that is ready for PCR.

Running PCR: Cycle 1 for the Pre-Denaturation process, Cycle 2 for the process of denaturation, annealing, extension. Cycle 3 for the final extension process.

Gel Electrophoresis: A total of 8 μ l amplicons from the PCR (Positive Control, Negative Control, sample) were inserted into 10 μ l gel wells and in the first hole add 10 μ l of 100 bp DNA ladder into the well near the positive control. Electrophoresis is turned on and run from a negative charge (cathode) to a positive charge (anode) at 100 A and 60 minutes. After electrophoresis, the band is formed. The positive result if the band is aligned with the positive control, continued by DNA Sequence examination to obtain the nucleotide sequence which was then performed a BLAST (Basic Local Alignment Search Tool) Nucleotide sequence on NCBI Gene Bank data (National Center for Biotechnology Information)

RESULT





Figure 1: Electrophoresis results of PCR amplification of Granulicatella adiacens samples in periodontitis patients

The study used PCR targeting the 16S rRNA gene for the detection of bacterial DNA. From the PCR amplification results obtained DNA band fragments of 100 bp in all isolation samples. Then proceed with the DNA sequence

and the results obtained in the form of nucleotide fasta, which then processed the data based on the order of nucleotides in NCBI Gene Bank data, using the BLAST analysis.

Table 1: Blast NCBI analysis for Granulicatella adiancense

Sample	Tooth number	Pocket Periodontal Depth (mm)	Data NCBI
1	21	10	+
2	11	11	+
3	24	9	+
4	31	7	-

5	12	5	-
6	27	6	+
7	21	5	-
8	16	7	+
9	23	11	+
10	31	8	+
11	17	6	-
12	27	9	+
13	16	5	+
14	16	7	+
15	34	5	-
16	31	8	+
17	12	9	+
18	36	8	+

DNA sequence results in 18 gram-positive bacterial isolation samples obtained by the Nucleotide sequence were then adjusted to the NCBI Gene Bank so that 13 samples

were obtained with the DNA of the bacterium Granulicatella adiacens.

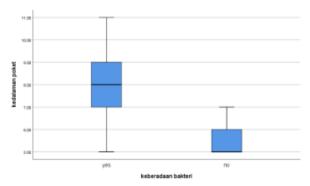


Figure 2: Granulicatella adiacens distribution based on periodontal depth

In figure 2 there were Granulicatella adiacens found on the periodontal pocket with 7-9 mm depth (SD±2mm), while no Granulicatella adiacens were found on 5 to 6 mm periodontal pocket depth (SD±1mm). From 18 bacterial isolation samples, 3 samples collect from pocket periodontal with more then 10 mm depth, 1 sample is collected from 6 mm depth and 9 samples from 7 to 9 mm pocket depth, all of this 13 samples were found obtained with the DNA of the bacterium Granulicatella adiacens. The remain samples is no Granulicatella adiacens found.

Based on the Kruskal-Wallis Test, the results showed that there is a difference between the depth of the pocket with a shallow pocket against the presence of the bacterium Granulicatella adiacens. Through observation, a significant difference (0.008) was obtained at different pocket depth levels, this difference was in the presence or absence of Granulicatella adiacens found at the depth of the periodontal pocket. From the results above that the bacterium Granulicatella adiacens was found at a depth of more than 7 mm.

DISCUSSION

The conventional PCR technique was chosen for bacterial identification because of its accuracy in checking the presence of bacterial bands, we also using DNA sequence methods, where this method aims to complete the examination of bacteria with Conventional PCR which results are used as a basis for nucleotide examination on the NCBI Gene Bank using BLAST analysis. In conventional PCR examination, a 16S rRNA universal primer is used, however, the data produced does not show the type of bacteria, but the quality of the bacteria through the presence of a bacterial band. On examination using the DNA sequence method, the type of bacteria can be clearly identified, both at the level of genus, species and even subspecies of bacteria.

The bacteria are the predominant group of organisms in the oral cavity, and there are probably some 500 to 700 common oral species or phylotypes which is only 50% to 60% are cultivable¹¹. In periodontitis condition, the discoveryof streptococcus species and their variants in large numbers, this indicates the possibility of a shift in bacterial species from healthy periodontal to periodontitis¹². Hajishengallis

(2013) revealed that periodontal disease is no longer solely caused by the presence of specific bacteria but by ecological changes in the biofilm community and bacterial species population level. *Colonization by the main pathogen is aided by additional pathogens causing impaired host defense and increasing inflammation 13.

Based on the results of the study it was found that there was a relationship between the depth of the periodontal pocket with the Granulicatella adiacens bacteria, where the deeper the periodontal pocket the bacteria could be found. This is consistent with bacterial morphology in the form of small Gram-positive, non-motile, non-spore coccus, oxidase, and catalase-negative, facultative anaerobes 14. The anaerobic traits of the facultative bacterial causes these bacteria to develop in deep periodontal pockets, in additional it was bacteria in the phylum firmicutes that classified into lactic acid bacteria, commensal bacteria in the oral cavity. Which has the ability of demineralization and bone resorption 15

Granulicatella adiacens is part of a normal microbiota in the oral cavity. urogenital. and intestinal tract. Protein secretion can help bacteria in normal growth physiology, otherwise, it has a function as a virulence mechanism in host colonization or by modulating host immune response.16 There are more than 20 putative virulence- related proteins for Granulicatella adiacens survival and cause infection, bacterial colonization, and virulence17. Protein secretion helps bacteria in their normal growth and also function as a virulence mechanism in host colonization or by modulating host immune Bacteria have devised dedicated secretory systems for protein secretion¹⁹, Gram-positive species secrete mainly via general secretory system²⁰. Virulence potential of the Granulicatella adiacens secretome assessed by searching for their associations with virulence activities in other species. Only a view study is available for the protein secretion of Granulicatella Recently, a close phylogenetic relative of adiacens. Granulicatella elegans, was shown to secrete arginine deiminase, which through citrullination inhibits proliferation of human peripheral blood mononuclear cells in vitro and may associate with the pathogenesis of periodontitis and certain systemic1621.

Putative virulence-associated proteins in Granulicatella adiacens secretome such as serine protease, thioredoxin, superoxide dismutase, phosphor carrier, and acyl carrier proteins²². Superoxide dismutase converts superoxide anions into oxygen and hydrogen peroxide, it is required not only for H₂O₂ resistance in S. mutans, but also is needed for coexistence with S. sanguinis23. Serine proteases cause cytopathic effects and exhibit enterotoxin activity, it degrades mucins, resulting in an advantage for mucosal colonization and immune modulation²⁴. In Salmonella enterica, thioredoxin helps the bacterium in intracellular virulence 2027 2009. Granulicatella replication and adiacens secretome consisted of several moonlighting proteins shown to play a role in bacterial virulence. The glycolytic enzymes, phosphoglycerate kinase, triosephosphate isomerase, aldolase, and enolase possess moonlighting properties, they function as adhesins,

receptors for transferrin, neutrophil evasion proteins immunomodulators and participate in extracellular polysaccharide synthesis. Oral bacteria express a number of molecular chaperones found in Granulicatella adiacens secretome. They express on the cell surface to use them as adhesins and can release them into the extracellular milieu to act as signaling virulence factors¹⁷.

CONCLUSION

Granulicatella adiancens is a normal flora bacteria in the oral cavity and in chronic periodontitis conditions, the presence of this bacteria is detected. There was a relationship between the depth of the periodontal pocket with the Granulicatella adiacens bacteria, where the deeper of the periodontal pocket the bacteria could be found.

ACKNOWLEDGMENT

The author is very grateful to IDTA member, especially to Adytha, Raodah Ramadhani and Jennifer Tjokro for the advices and discussions

CONFLICT OF INTERESTS

There are no conflict of interests in this study

REFERENCES

- IGAAP S, Manuaba IP, Thahir H, et al. (2019). The Effectiveness Of Giving Snail Slime (Acatina Fulica) On The Healing Of Pocket On The Wistar Rats With Periodontitis. Int J Appl Pharm. 11(4):1-3. doi:http://dx.doi.org/10.22159/ijap.2019.v11s4.352
 - doi:http://dx.doi.org/10.22159/ijap.2019.v11s4.352 81
- Dharmawati IGAAYU, Manuaba IBP, Thahir H, et al. (2019). Pocket Measurement Methods In Wistar Rats Induced by Bacteria and The Instalation Of Silk Ligature: An Experimental Studies. Int J Appl Pharm.11(4):1-4. doi:http://dx.doi.org/10.22159/ijap.2019.v11s4.352 96
- Kapoor A, Malhotra R, Grover V, Grover D. (2012). Systemic Antibiotic Therapy In Periodontics. Dent Res J (Isfahan). 9(5):505-515.
- Hajishengallis G, Lamont RJ. (2013). Beyond The Red Complex and Into More Complexity: The Polymicrobial Synergy and Dysbiosis (PSD) Model of Periodontal Disease Etiology. Mol Oral Microbiol. 27(6):409-419. doi:10.1111/j.2041-1014.2012.00663.x.Beyond
- Bowen WH, Burne RA, Wu H, Koo H. (2017).
 Oral Biofilms: Pathogens, Matrix, and Polymicrobial Interactions in Microenvironments.
 Trends Microbiol. 1:1-14. doi:10.1016/j.tim.2017.09.008
- Christensen JJ, Facklam RR. (2001). Granulicatella and Abiotrophia Species from Human Clinical Specimens. J Clin Microbiol. 39(10):3520-3523. doi:10.1128/JCM.39.10.3520
- Karched M, Bhardwaj RG, Asikainen SE. (2015).
 Coaggregation and Biofilm Growth of

- Granulicatella spp. With Fusobacterium nucleatum and Aggregatibacter actinomycetemcomitans. BMC Microbiol. 15(114):1-10. doi:10.1186/s12866-015-0439-z
- Cargill JS, Scott KS, Gascoyne-binzi D, Sandoe JAT. (2012). Granulicatella Infection: Diagnosis and Management. J Med Microbiol. 61:755-761. doi:10.1099/jmm.0.039693-0
- Ling Y, Su Q, Jiang S, Wu Q, Qian Y, Hospital N. (2016). Granulicatella adiacens: an Unusual Cause of Early-Onset Pneumonia in a Neonate. Clin Microbiol Newsl. 38(3):24-25. doi:10.1016/j.clinmicnews.2016.01.002
- Cerceo E, Christie JD, Nachamkin I, Lautenbach E. (2004). Central Nervous System Infections due to Abiotrophia and Granulicatella species: an Emerging Challenge? Diagn Microbiol Infect Dis. 48:161-165. doi:10.1016/j.diagmicrobio.2003.10.009
- Samaranayake L, Matsubara VH. (2017). Normal Oral Flora and the Oral Ecosystem. Dent Clin N Am. 61:199-215. doi:10.1016/j.cden.2016.11.002
- Lamont RJ, Hajishengallis G. (2016).
 Polymicrobial Synergy and Dysbiosis in Inflammatory Disease. Trends Mol Med. 21(3):172-183.
- doi:10.1016/j.molmed.2014.11.004.Polymicrobial

 13. Lamont RJ, Koo H, Hajishengallis G. (2018). The
- Oral Microbiota: Dynamic Communities and Host Interactions. Nat Rev Microbiol. 16:745-759. doi:10.1038/s41579-018-0089-x
- Ruoff KL. (1991). Nutritionally Variant Streptococci. Clin Microbiol Rev. 4(2):184-190.
- Hajishengallis G, Liang S, Payne MA, et al. (2011).
 A Low-Abundance Biofilm Species Orchestrates Inflammatory Periodontal Disease through the Commensal Microbiota and the Complement Pathway. Cell Host Microbe. 10(5):497-506. doi:10.1016/j.chom.2011.10.006.A
- Olsen I, Singhrao SK, Potempa J. (2018).
 Citrullination as a Plausible Link to Periodontitis,
 Rheumatoid Arthritis, Atherosclerosis and
 Alzheimer's Disease. J Oral Microbiol. 10(1):1-8.
 doi:10.1080/20002297.2018.1487742
- Karched M, Bhardwaj RG, Tiss A, Asikainen S. (2019). Proteomic Analysis and Virulence Assessment of Granulicatella adiacens Secretome. Front Cell Infect Microbiol. 9:1-12. doi:10.3389/fcimb.2019.00104
- Lee VT, Schneewind O. (2001). Protein Secretion and The Pathogenesis of Bacterial Infections. Genes Dev. 15:1725-1752. doi:10.1101/gad.896801
- GreenER, Mecsas J. (2016). Bacterial Secretion Systems: An Overview. Microbiol Spectr. 4(1):1-19. doi:10.1128/microbiolspec.VMBF-0012-2015.
- Schneewind O, Missiakas DM. (2012). Protein Secretion and Surface Display in Gram-positive Bacteria. Phil Trans R Soc B. 367:1123-1139. doi:10.1098/rstb.2011.0210
- 21. Kanamoto T, Sato S, Nakashima H, Inoue M.

- (2007). Proliferation of Mitogen-Stimulated Human Peripheral Blood Mononuclear Cells is Inhibited by Extracellular Arginine Deiminase of Granulicatella elegans Isolated From the Human Mouth. J Infect Chemother.13:353-355. doi:10.1007/s10156-007-0546-3.
- 22. Ramadhany S, Achmad H, Handayani H, Tanumihardja M, Singgih MF, Inayah NH, Ramadhany YF. (2020). Formulation of Ethanol Extract (Myrmecodiapendans) as an Antibacterial Streptococcus mutansin Chewable Lozenges for Children with Early Childhood Caries. Sys Rev Pharm. 11(4): 252 257.
- Stobernack T, Glasner C, Junker S, et al. (2016).
 The Extracellular Proteome and Citrullinome of the Oral Pathogen Porphyromonas gingivalis The Extracellular Proteome and Citrullinome of the Oral Pathogen Department of Medical Microbiology, University of Groningen, University Medical Ernst-Moritz-Arndt. J Proteome Res. 1-44. doi:10.1021/acs.jproteome.6b00634.
- Achmad H, Tanumihardja M, Sartini, Ramadhany S, Singgih MF, Ramadhany YF, Mutmainnah N. (2020). Chewable Lozenges using White Shrimp Waste (Litopenaeusvannamei) in Reduce Colonization of Bacteria Streptococcus mutansin the Case of Early Childhood Caries. Sys Rev Pharm. 11(4): 293 299.
- Fujishima K, Kawada-matsuo M, Oogai Y, Tokuda M, Torii M. (2013). dpr and sodIn Streptococcus mutans Are Involved in Coexistence With S. sanguinis, and PerR Is Associated with Resistance to H₂O₂. Appl Environ Microbiol. 79(5):1436-1443. doi:10.1128/AEM.03306-12
- Achmad H, Thahir H, Rieuwpassa I, Mardiana AA, Oktawati S, Samad R, Djais AI, Gani A, Singgih MF, Madjid F, Admy SC. (2020). The Effectiveness of Channastriata ExtractAntimicrobial Effect on Periopathogen Bacteria (Porphyromonasgingivalis and Aggregatibacteractinomycetemcomitans). Sys Rev Pharm. 11(4): 319 323
- Ruiz-Perez F, James N. (2013). Bacterial Serine Proteases Secreted by The Autotransporter Pathway: Classification, Specificity, and Role in Virulence. Cell Mol Life Sci. 71(5):745-770. doi:10.1007/s00018-013-1355-8
- Bjur E, Eriksson-ygberg S, Fredrik A, Rhen M. (2006). Thioredoxin 1 Promotes Intracellular Replication and Virulence of Salmonella enterica Serovar Typhimurium. Infect Immun. 74(9):5140-5151. doi:10.1128/IAI.00449-06.
- Achmad H, Adam AM, Mappangara S, Oktawati S, Sjahril R, Singgih MF, Neormansyah I, Siswanto H. (2020). Identification and Antimicrobial Susceptibility of GranulicatellaadiacensIsolated from Periodontal Pocket. Sys Rev Pharm. 11(4): 324 331.

Granulicatella Adiacens Bacteria Isolation from Perodontitical Patients with Polymerase Chain Reaction Techniques

ORIGINALITY REPORT INTERNET SOURCES **PUBLICATIONS** SIMILARITY INDEX STUDENT PAPERS **PRIMARY SOURCES** www.frontiersin.org **6**% Internet Source Submitted to Udayana University Student Paper Submitted to Universitas Katolik Indonesia Atma Jaya Student Paper sysrevpharm.org Internet Source Maribasappa Karched, Radhika G. Bhardwaj, Ali Tiss, Sirkka Asikainen. "Proteomic Analysis and Virulence Assessment of Granulicatella adiacens Secretome", Frontiers in Cellular and Infection Microbiology, 2019 Publication link.springer.com Internet Source

7 archive.org

8

worldwidescience.org

Internet Source

<1%

9

Vernal, Rolando, Alejandra Chaparro, Rebecca Graumann, Javier Puente, Maria Antonieta Valenzuela, and Jorge Gamonal. "Levels of Cytokine Receptor Activator of Nuclear Factor kB Ligand in Gingival Crevicular Fluid in Untreated Chronic Periodontitis Patients", Journal of Periodontology, 2004.

• /(

Publication

Exclude quotes

Off

Exclude matches

Off

Exclude bibliography

On