ci,_heparan_sulfate,_and_synde can_expression_in_HUVECs_cell s.pdf

Submission date: 01-Mar-2022 08:26AM (UTC+0700)

Submission ID: 1773411345

File name: ci,_heparan_sulfate,_and_syndecan_expression_in_HUVECs_cells.pdf (1.68M)

Word count: 7822

Character count: 43125

Research Article

Effects of single and intermittent glucose exposure on hyaluronic acid, heparan sulfate, and syndecan expression in HUVECs cells

DEWI INDAH NOVIANA PRATIWI^{1,2*}, NIA KANIA³, M ARIS WIDODO⁴, KUSWORINI HANDONO⁵

¹Doctoral Program in Medicine, Faculty of Medicine, Universitas Brawijaya, Malang, East Java, Indonesia

²Department of Clinical Pathology, Faculty of Medicine, Universitas Lambung Mangkurat/Ulin General Hospital, Banjarmasin, South Kalimantan, Indonesia

³Department of Pathology, Faculty of Medicine, Universitas Lambung Mangkurat/Ulin General Hospital, Banjarmasin, South Kalimantan, Indonesia

⁴Department of Pharmacology, Faculty of Medicine, Universitas Brawijaya, Malang, East Java, Indonesia

⁵Department of Clinical Pathology, Faculty of Medicine, Universitas Brawijaya/Dr. Saiful Anwar General Hospital, Malang, East Java, Indonesia

*Corresponding author:

Email: indahhariadi@gmail.com*

Received: 20.04.20, Revised: 17.05.20, Accepted:03.06.20

ABSTRACT

Objectives: This study aims to investigate 1) the effect of incubation duration on changes in HUVECs cell glycocalyx expression, 2) the impact of glucose dose on changes in HUVECs cell glycocalyx expression, and 3) the comparison of single and intermittent glucose doses on changes in HUVECs cell glycocalyx expression.

Methods: After confluence, the HUVECs were divided into three steps to study. On the first steps, HUVECs were divided into control and duration of incubation of 12-hours; 24-hours; 48-hours and 72-hours. On the second step, HUVECs were classified according to the glucose concentration exposure (0 mM; 5 mM or 22 mM) for 72-hours of incubation. On the third step. HUVECs were divided into single and intermittent doses (for 72-hours of incubation).

Results: The expression of hyaluronic acid, heparan sulfate, and syndecan was analyzed by laser scanning confocal microscope.

Conclusions: In conditions without glucose exposure and physiological glucose exposure, glycocalyx has a dynamic expression. High glucose exposure eliminates the dynamics of hyaluronic acid and syndecan expression. Single-dose exposure triggers a decrease in hyaluronic acid expression compared to intermittent exposure, and this is the opposite of heparan sulfate exposure.

Keywords: glycocalyx, endothelial cells, single dose, intermittent dose.

INTRODUCTION

Endothelial cells are complex endocrine and paracrine organs that play a physiological role in the modulation of vascular tone (vasoconstriction and vasodilation), hemostasis, growth regulation and differentiation of vascular smooth muscle cells, and inflammatory modulation (Hossain, Kawar and El Nahas, 2007). Diabetes mellitus is a chronic endocrine disease due to relative or absolute insulin deficiency (Kristensen et al., 2016). Cardiovascular complications are a life threat to people with diabetes mellitus, and their progress is mostly determined by the status of hyperglycemia (Dalan et al., 2014; Fox et al.,

2015). Hyperglycemia triggers dysfunction, damage, and death of endothelial cells and acceleration of atherosclerosis as a basis for cardiovascular complications (Deguchi and Miyazaki, 2010; Georgescu et al., 2011; Bammert et al., 2017).

Endothelial cell function is determined by the structure of the cytoskeleton and glycocalyx (Fels et al., 2014). Glycocalyx consists of glycosaminoglycans, proteoglycans, and glycoproteins (Barakat, 2008; Fu and Tarbell, 2013; Sieve, Münster-Kühnel and Hilfiker-Kleiner, 2018). Hyaluronic acid is a linear glycosaminoglycan that is chemically composed

of N-acetylglucosamine and D-glucoronic acid through b-1.3 alycosidic bonds and disaccharide units which are bound to b-1.4 glycosidic (Huang, Chen and Chen, 2018). Heparan sulfate is linear polysaccharides arranged by N-acetylated or Nsulfonated glucosamine units and uronic acid. Heparan sulfate occupy 50% -90% glycosaminoglycan (Tarbell and Pahakis, 2006; Reitsma et al., 2007; Saito, 2015). Syndecan-1 transmembrane is a core protein of glycocalyx. Hyaluronic acid chains and heparan sulfate attach to syndecan-1 and are located on the lumen surface of the glycocalyx network (Saito, 2015).

things underlie the emergence of complications of diabetes mellitus, the degree of hyperglycemia, and hyperglycemia fluctuations (Home, 2005). Acute hyperglycemia triggers glycocalyx degradation. Exposure to high glucose levels triggers syndecan shedding. Diabetes mellitus triggers an increase in hyaluronic acid plasma as well as shedding syndecan (Nieuwdorp et al., 2006; Broekhuizen et al., 2010). Glucose fluctuations (peak hyperglycemia hypoglycemia) will trigger the production of reactive oxygen compounds and cellular apoptosis, which are more massive than persistent high sugar levels (Ricks et al., 2012; Quincozes-Santos et al., 2017). In HUVECs cells exposed to glucose 5.5 mM and 22 mM alternatively, reactive oxygen compounds will be formed, which are more significant than constant exposure (Quincozes-Santos et al., 2017). To the best of our knowledge, the effects of hyperglycemia fluctuations on glycocalyx are unclear. Therefore, this study aims to investigate 1) the impact of incubation duration on changes in HUVECs cell glycocalyx expression, 2) the effect of glucose dose on changes in HUVECs cell glycocalyx expression, and 3) comparison of single and intermittent glucose doses on changes in HUVECs cell glycocalyx expression.

Methods

Multure cells

HUVEC cells were purchased from ATCC. The cells were cultured in RPMI-1640 medium supplemented (10% fetal bovine serum, 100 U/mL penicillin-streptomycin), and cultured at 37°C in 5% CO2 incubator. The cells were passaged every 3 to 4 days, with 0.25% trypsin

(w). The HUVEC cells in the logarithmic growth phase were inoculated in 96-well plate at a density of 1 \times 105 cells/mL, 100 $\mu\text{L/well}$. The cells were incubated at 37°C, 5% CO2 in a cell incubator for 24 h, and the original culture medium was replaced with a fresh medium, and the cells were divided into three steps of the study. The first steps were to explore the effect of incubation duration on changes in HUVECs cell glycocalyx expression. The second steps were to investigate the impact of glucose dose on changes in HUVECs cell glycocalyx expression. The third step was to analyze the comparison of single and intermittent glucose doses on changes in HUVECs cell glycocalyx expression.

Immunofluorescence

Immunofluorescence analysis was carried out according to the procedure in previous studies (Khanmohammadi, Sakai and Taya, 2017). The expression of glycocalyx were detected by the primary antibody of hyaluronic acid (Santa Cruz Biotechnology Inc, Dallas, Texas, USA, Catalog number Sc-221733), heparan sulfate (Antibodies Online, Aachen, Germany, Catalog number ABIN2280658), and syndecan (Bioss Antibodies Inc, Woburn, Massachusetts, USA, Catalog number Bs-1309R-Cy5).

Statistical analysis

Data was presented in mean ± standard deviation. Data were analyzed by ANOVA test using SPSS version 16 for Windows.

Results

Effect of incubation duration on glycocalyx

Figure 1 presents the expression of hyaluronic acid in HUVECs cells. The hyaluronic acid expression tends to increase in 24-hour incubation compared to 12-hour incubation, although it has not been significantly different (p > 0.05). Hyaluronic acid expression was significantly lower at 48-hour incubation compared to 12-hour incubation (p < 0.05). Hyaluronic acid expression was significantly higher at 72-hour incubation compared to 12 hours, 24 hours, or 48-hour incubation (p < 0.05). Hyaluronic acid expression was not significantly different at 48-hour incubation compared to 24-hour incubation (p > 0.05).

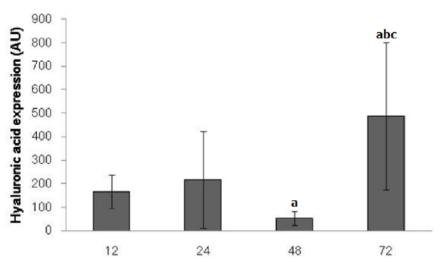


Fig.1. The expression of hyaluronic acid according to incubation time (without glucose exposure).

Note: data was pre interest as mean ± standart of deviation; a: p < 0.05 in comparison with 12-hours of incubation. b: p < 0.05 in comparison with 24-hours of incubation. c: p < 0.05 in comparison with 48-hours of incubation. AU: arbitrary units.

Heparan sulfate expression in HUVECs cells can be seen in Figure 2. Heparan sulfate expression significantly increased at 24-hour, 48-hour incubation compared to 12-hour incubation (p < 0.05). Heparan sulfate expression did not differ significantly at 72-hour incubation compared to 12-hour incubation (p > 0.05). Heparan sulfate

expression did not differ significantly at 48 hours or 72 hours incubation compared to 24 hours incubation (p > 0.05). Heparan sulfate expression did not differ significantly at 72 hours incubation compared to 48 hours incubation (p > 0.05).

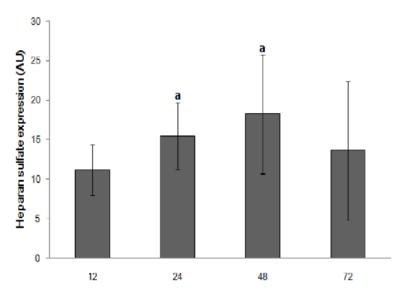


Fig. 2:The expression of heparan sulfate according to incubation time (without glucose exposure). Note: data was presented as mean \pm standart of deviation; a: p < 0.05 in comparison with 12-hours of incubation. AU: arbitrary units.

Figure 3 displays the syndecan expression of HUVECs cells in various incubation groups. Syndecan expression was significantly lower at 24-hour, 48-hour, and 72-hour incubation compared to 12-hour incubation (p < 0.05).

Syndecan expression did not differ significantly at 48 hours or 72 hours incubation compared to 24 hours incubation (p > 0.05). Syndecan expression did not differ significantly at 72 hours incubation compared to 48 hours incubation (p > 0.05).

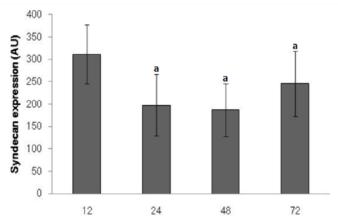


Fig.3:The expression of syndecan according to incubation time (without glucose exposure). Note: data was presented as mean ± standart of deviation; ^a: p < 0.05 in comparison with 12-hours of incubation. AU: arbitrary units.

Figure 4 presents the expression of hyaluronic acid in HUVECs exposed to 5 mM glucose. Hyaluronic acid expression was significantly higher at 24-hour, 48-hour, and 72-hour incubation compared to 12-hour incubation (p < 0.05). Hyaluronic acid expression was significantly lower at 48-hour incubation than 24-

hour incubation (p < 0.05). The Hyaluronic acid expression did not differ significantly at 72-hour incubation compared to 24-hour incubation (p < 0.05). Hyaluronic acid expression was significantly higher at 72 hours incubation than 48 hours incubation (p < 0.05).

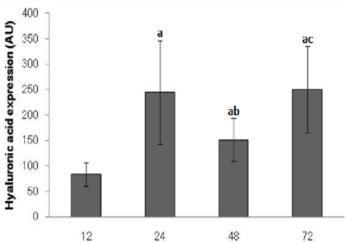


Fig.4. The expression of hyaluronic acid according to cubation time (glucose 5 mM). Note: data was preserted as mean ± standart of deviation; a: p < 0.05 in comparison with 12-hours of incubation. b: p < 0.05 in comparison with 24-hours of incubation. c: p < 0.05 in comparison with 48-hours of incubation. AU: arbitrary units.

Heparan sulfate expression at five mM glucose exposure to HUVECs cells can be seen in Figure 5. Heparan sulfate expression did not differ significantly at 24-hour, 48-hour incubation compared with 12-hour incubation (p < 0.05). Heparan sulfate expression significantly decreased at 72-hour incubation compared to

12-hour incubation (p < 0.05). Heparan sulfate expression significantly decreased at 48-hour incubation compared to 24-hour incubation (p > 0.05). Heparan sulfate expression was significantly lower at 72-hour incubation than 24-hour incubation (p > 0.05).

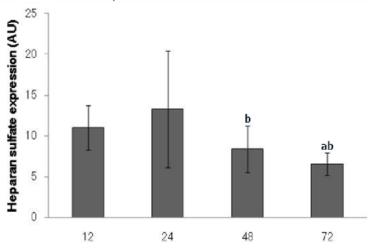


Fig.5. The expression of heparan sulfate according to cubation time (glucose 5 mM). Note: data was presented mean ± standart of deviation; 2: p < 0.05 in comparison with 12-hours of incubation. b: p < 0.05 in comparison with 24-hours of incubation. AU: arbitrary units.

Figure 6 shows the change in syndecan expression in HUVECs exposed to 5 mM glucose. Syndecan expression significantly increased at 24-hour, 48-hour, and 72-hour incubation compared to 12-hour incubation (p < 0.05). Syndecan expression was significantly higher at

incubation 48 compared to 24-hour incubation (p < 0.05). Syndecan expression did not differ significantly at 72-hour incubation compared to 24-hour incubation (p > 0.05). Syndecan expression was significantly lower at 72 hours incubation than 48 hours incubation (p < 0.05).

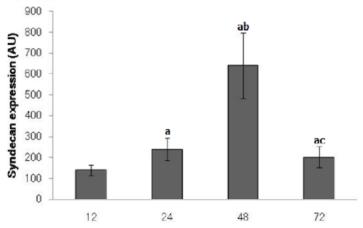


Fig.6:The expression of syndecan according incubation time (glucose 5 mM). Note: data was presented as mean ± standart of deviation; a: p < 0.05 in comparison with 12-hours of incubation. b: p < 0.05 in comparison with 24-hours of incubation. c: p < 0.05 in comparison with 48-hours of incubation. AU: arbitrary units.

Hyaluronic acid expression due to high-dose glucose exposure is presented in Figure 7. Hyaluronic acid expression was significantly higher at 24, 48 hours, and 72 hours incubation than 12 hours incubation (p < 0.05). Hyaluronic acid expression was not significantly different at 48-hour incubation compared to 24-hour

incubation (p < 0.05). Hyaluronic acid expression significantly decreased at 72-hour incubation compared to 24-hour incubation (p > 0.05). Hyaluronic acid expression significantly decreased at 72 hours incubation compared to 48 hours incubation (p > 0.05).

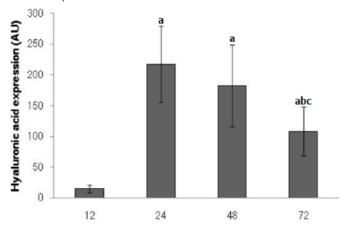


Fig.7:The expression of hyaluronic acid according to is ubation time (glucose 22 mM). Note: data was presersed as mean ± standart of deviation; a: p < 0.05 in comparison with 12-hours of incubation. b: p < 0.05 in comparison with 24-hours of incubation. c: p < 0.05 in comparison with 48-hours of incubation. AU: arbitrary units.

Figure 8 displays the expression of heparan sulfate due to high doses of glucose. Heparan sulfate expression did not differ significantly at 24-hour incubation compared with 12-hour incubation (p > 0.05). Heparan sulfate expression significantly decreased at 48-hour incubation compared to 12-hour incubation (p > 0.05). Heparan sulfate expression did not differ significantly at 72-hour incubation compared to

12-hour incubation (p > 0.05). Heparan sulfate expression was significantly lower at 48-hour incubation than 24-hour incubation (p > 0.05). Heparan sulfate expression did not differ significantly at 72-hour incubation compared to 24-hour incubation (p > 0.05). Heparan sulfate expression was significantly increased in 72-hour incubation compared to 48-hour incubation (p > 0.05).

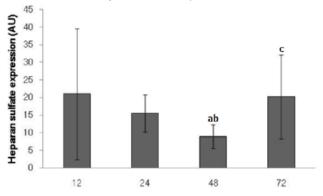


Fig.8:The expression of heparan sulfate according to Eubation time (glucose 22 mM). Note: data was preser data was preser and as mean ± standart of deviation; a: p < 0.05 in comparison with 12-hours of incubation. b: p < 0.05 in comparison with 24-hours of incubation. c: p < 0.05 in comparison with 48-hours of incubation. AU: arbitrary units.

Syndecan expression due to high-dose glucose exposure can be seen in Figure 9. Syndecan expression was not significantly different at 24-hour incubation than 12-hour incubation (p < 0.05). Syndecan expression significantly increased at 48 hours or 72 hours incubation compared to

12 hours incubation (p < 0.05). Syndecan expression significantly increased at 48-hour incubation compared to 24-hour incubation (p < 0.05). Syndecan expression significantly increased at 72-hour incubation compared to 24-hour or 48-hour incubation (p < 0.05).

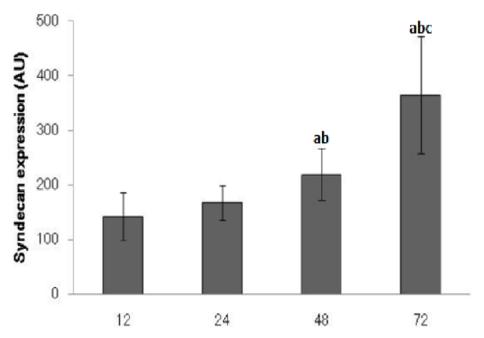


Fig.9:The expression of syndecan according 3 incubation time (glucose 22 mM). Note: data was pr3 ented as mean ± standart of deviation; a: p < 0.05 in comparison with 12-hours of incubation. b: p < 0.05 in comparison with 24-hours of incubation. c: p < 0.05 in comparison with 48-hours of incubation. AU: arbitrary units.

Effect of glucose dosage on glycocalyx

Figure 10 displays the expression of hyaluronic acid HUVECs cells exposed to various doses of glucose in different incubation periods. In 12-hour incubation, the expression of hyaluronic acid decreased significantly at a dose of 5 mM or 22 mM compared to a glucose dose of 0 mM (p < 0.05). Hyaluronic acid expression significantly decreased at 22 mM dose compared to a 5 mM glucose dose (p < 0.05). In 24-hour incubation, the expression of hyaluronic acid did not differ significantly at a dose of 5 mM compared to a dose of 0 mM glucose or a dose of 22 mM compared to a dose of 0 mM glucose (p < 0.05).

At 48 hours incubation, the expression of hyaluronic acid was significantly higher at a dose of 5 mM or 22 mM than a glucose dose of 0 mM (p < 0.05). Hyaluronic acid expression was not significantly different at 22 mM compared to 5 mM glucose dose (p > 0.05). At 72 hours incubation, the expression of hyaluronic acid did not differ significantly at a dose of 5 mM compared to a glucose dose of 0 mM (p > 0.05). Hyaluronic acid expression significantly decreased at a dose of 22 mM compared to a glucose dose of 0 mM (p < 0.05). Hyaluronic acid expression significantly decreased at 22 mM dose compared to a 5 mM glucose dose (p < 0.05).

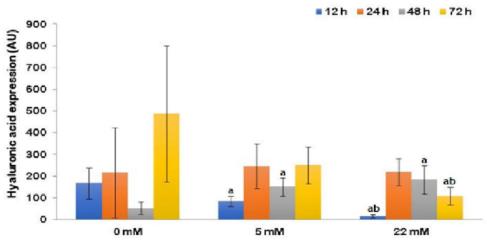


Fig.10:The expression of hyalur spic acid according to glucose level. Note: data was presented spic acid according to glucose level. Note: data was presented spice mean ± standart of deviation; a: p < 0.05 in comparison with glucose level at dose of 0 mM. b: p < 0.05 in comparison with glucose level at dose of 22 mM. AU: arbitrary units.

Heparan sulfate expression of HUVECs cells exposed to various doses of glucose across multiple incubation periods can be seen in Figure 11. At 12 hours, 24 hours, and 72 hours incubation, heparan sulfate expression did not differ significantly at five mM or 22 mM doses compared to 0 mM or between a dose of 22 mM

compared to a dose of 5 mM (p > 0.05). At 48 hours incubation, heparan sulfate expression significantly decreased at a dose of 5 mM or 22 mM compared to a glucose dose of 0 mM (p < 0.05). Heparan sulfate expression did not differ significantly at a dose of 22 mM compared to a glucose dose of 5 mM (p > 0.05).

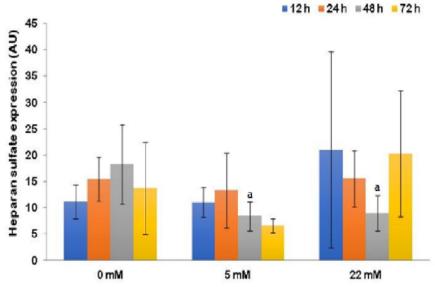


Fig.11:The expression of heparan sulfate according to glucose level. Note: data was presented as mean \pm standart of deviation; ^a: p < 0.05 in comparison with glucose level at dose of 0 mM. AU: arbitrary units.

Figure 12 shows the syndecan expression of HUVECs cells exposed to various doses of glucose in various incubation periods. At 12-hour incubation, syndecan expression was significantly lower at a dose of 5 mM or a dose of 22 mM compared to 0 mM (p < 0.05). Syndecan expression did not differ significantly at a dose of 22 mM compared to a dose of 5 mM (p > 0.05). At 24-hour incubation, syndecan expression was not significantly different at a dose of 5 mM compared to a dose of 0 mM glucose or a dose of 22 mM compared to a dose of 0 mM glucose or a dose of 22 mM compared to a dose of 0 mM glucose (p < 0.05). At 48 hours incubation, syndecan expression was significantly higher at five mM

dose than glucose dose of 0 mM (p < 0.05). Syndecan expression did not differ significantly at a dose of 22 mM compared to a glucose dose of 0 mM (p < 0.05). Syndecan expression significantly decreased at 22 mM dose compared to 5 mM glucose dose (p < 0.05). At 72 hours incubation, syndecan expression did not differ significantly at a dose of 5 mM compared to a glucose dose of 0 mM (p > 0.05). Syndecan expression significantly increased at 22 mM compared to glucose 0 mM (p < 0.05). Syndecan expression was significantly higher at 22 mM dose than a five mM glucose dose (p < 0.05).

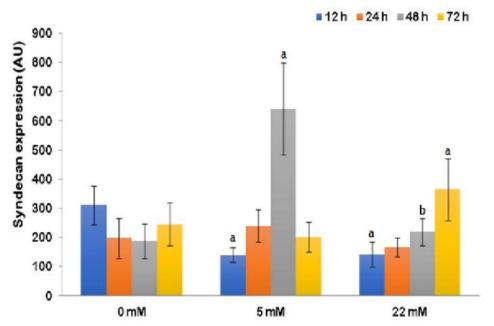


Fig.12:The expression of 5 ndecan according to glucose level. Note: data was presentes as mean ± standart of deviation; a: p < 0.05 in comparison with glucose level at dose of 0 mM. b: p < 0.05 in comparison with glucose level at dose of 22 mM. AU: arbitrary units.

Comparison of single and intermittent doses

Hyaluronic acid expression in various study groups can be seen in Figure 13. Hyaluronic acid expression was not significantly different in glucose five dose exposure compared with 0 mM glucose dose (p > 0.05). Hyaluronic acid expression significantly decreased at exposure to glucose dose 22 compared to glucose dose of 0 mM or intermittent treatment compared to

glucose dose of 0 mM (p < 0.05). Hyaluronic acid expression was significantly lower in glucose exposure at dose 22 compared with glucose dose of 5 mM (p > 0.05). Hyaluronic acid expression was not significantly different in intermittent glucose exposure compared to a 5 mM glucose dose (p > 0.05). Hyaluronic acid expression was significantly higher in glucose occasional dose exposure than 22 mM glucose dose (p > 0.05).

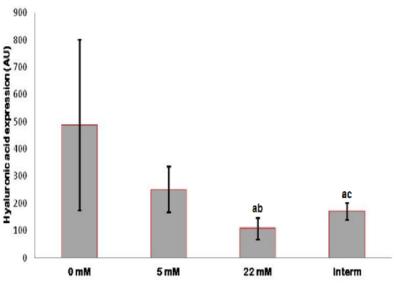


Fig. 13:The expression of hyaluronic acid in sing 5 and intermittent glucose exposure. Note: data was presente 5 as mean ± standart of deviation; a: p < 0.05 in comparison 5th glucose level at dose of 0 mM. b: p < 0.05 in comparison with glucose level at dose of 5 mM. c: p < 0.05 in comparison with glucose level at dose of 22 mM. AU: arbitrary units.

Heparan sulfate expression in various study groups can be seen in Figure 14. Heparan sulfate expression was significantly lower in glucose five dose exposure compared to 0 mM glucose dose (p < 0.05). Heparan sulfate expression did not differ significantly in glucose dose 22 compared with glucose dose of 0 mM (p > 0.05). Heparan

sulfate expression increased significantly in glucose exposure at dose 22 compared to glucose dose of 5 mM (p < 0.05). Heparan sulfate expression significantly increased with intermittent glucose exposure compared with five mM or 22 mM glucose (p < 0.05).

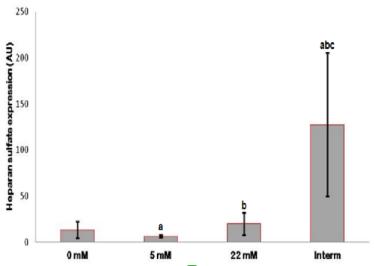


Fig.14:The expression of heparan sulfate in sing s and intermittent glucose exposure. Note: data was presentes as mean ± standart of deviation; a: p < 0.05 in comparison (5th glucose level at dose of 0 mM. b: p < 0.05 in comparison with glucose level at dose of 5 mM. c: p < 0.05 in comparison with glucose level at dose of 22 mM. AU: arbitrary units.

Syndecan expression in various groups can be seen in Figure 15. Syndecan expression was not significantly different in glucose five dose exposure compared with 0 mM glucose dose (p > 0.05). Syndecan expression significantly increased by exposure to glucose 22 doses compared to 0 mM glucose doses (p < 0.05). Syndecan expression did not differ significantly in intermittent glucose exposure compared with 0

mM glucose dose (p > 0.05). Syndecan expression significantly increased with exposure to glucose 22 doses compared to 5 mM glucose doses (p < 0.05). Syndecan expression did not differ significantly in intermittent glucose exposure compared to 5 mM glucose dose (p > 0.05). Syndecan expression significantly decreased in intermittent glucose exposure compared to 22 mM glucose dose (p < 0.05).

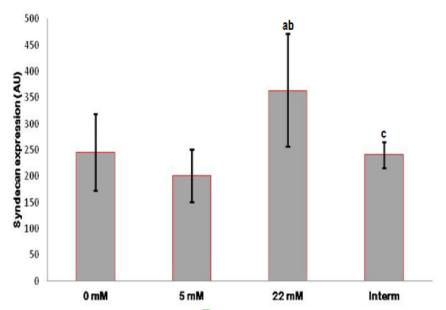


Fig.15:The expression of syndecan in single 3nd intermittent glucose exposure. Note: data was present 5 as mean ± standart of deviation; a: p < 0.05 in comparison 5th glucose level at dose of 0 mM. b: p < 0.05 in comparison with glucose level at dose of 5 mM. c: p < 0.05 in comparison with glucose level at dose of 22 mM. AU: arbitrary units.

DISCUSSION

Effect of incubation duration on glycocalyx

In this study, it was proven that hyaluronic acid, heparan sulfate, and syndecan-1 are present on the surface and expressed by HUVECs cells. This finding is consistent with previous findings that glycoprotein and proteoglycans are expressed in HUVECs (Savery et al., 2013; Jing et al., 2016; Li et al., 2016; Khanmohammadi, Sakai and Taya, 2017; Yang et al., 2019).

In this study, the dynamics of hyduronic acid, heparan sulfate, and syndecan expression were found in HUVECs cells for various incubation periods (without glucose exposure). Hyduronic acid expression was significantly higher at 72-hour incubation compared to 12-hour, 24-hour, or 48-hour incubation. This indicates that at 72 hours of incubation, there is a tendency toward synthesis compared to the degradation of

hyaluronic acid. Hyaluronic acid is synthesized in the plasma membrane through the activation of hyaluronic acid synthase. Hyaluronic acid degradation occurs due to gradual local deterioration (Saegusa, Isaji and Kawarada, 2002; Leskova et al., 2019), or due to hyaluronidase released by endothelial cells (Chajara et al., 2000; Chowdhury et al., 2016). Various enzymes are also involved in the degradation of hyaluronic acid, including thrombin, cathepsin B, proteinase-3, and plasmin (Becker et al., 2015). This study uses M-199 medium containing magnesium. Hyaluronic acid synthase requires magnesium as a catalyst (DeAngelis, 1996).

In this study, the length of incubation did not affect the expression of heparan sulfate. This indicates that changes in the local environment during incubation do not modify the expression of

heparan sulfate exposure. Syndecan was found to be significantly reduced at 24, 48, and 72 hours incubation compared to 12 hours. We suspect that this mechanism of decline is caused by the activation of matrix metalloproteinase (Zeng et al., 2014). Magnesium contained in the M-199 medium will be used for hyaluronic acid synthase catalysts in HUVECs cell membranes. This will result in magnesium deficiency as long as incubation. In HUVECs with magnesium deficiency, there will be an increase in matrix metalloproteinase activity (Ferrè et al., 2010).

On physiological dose glucose exposure (5 mM), there was a significant increase in hyaluronic acid expression at 24-hour, 48-hour, and 72-hour incubation compared to 12-hour incubation. This indicates that glucose exposure at physiological doses is a stimulus to the upregulation of hyaluronic acid HUVECs cells. Hyaluronic acid is glycocalyx, which is synthesized in cell membranes, not in the Golgi apparatus. The synthesis of hyaluronic acid is not only through hyaluronan synthase in cell membranes but is also played by cytoplasmic enzymes that produce uridine-diphosphate (UDP) -glucose precursors (Viola et al., 2008; Gallo et al., 2019). For heparan sulfate, there was a significant decrease in 72-hour incubation compared to 12-hour incubation. The synthesis of heparan sulfate is played by sulfotransferase in the Golgi Apparatus, while degradation of heparan sulfate is played by sulfatase and heparanase (Patel, Pineda and Hoffman, 2017). This shows that exposure to glucose in physiological doses causes degradation of heparan sulfate. We suspect this degradation mechanism occurs through the formation of oxidants from glucose exposure. Monosaccharides undergo autooxidation to form superoxide radicals, hydroxyl radicals, hydrogen peroxide, and carbon-centered radicals (Wolff and Dean, 1987). Glycosaminoglycans can be damaged by the oxidant system (Han and Hiebert, 2013; Marini et al., 2018).

For syndecan, there is a dynamics of fluctuation in syndecan expression on physiological doses of glucose exposure. Syndecan release occurs due to several physiological agents, for example, thrombin, hyperosmolarity, sphingomyelinase, and ceramide (Fitzgerald et al., 2000). Syndecan release due to glucose in physiological levels is thought to be protective against endothelial cells (Wang et al., 2009). Syndecan ectodomains dissolved due to shedding are biologically active and capable of binding to the same ligand (Subramanian, Fitzgerald and Bernfield, 1997). For high doses of glucose exposure, there was a

For high doses of glucose exposure, there was a significant increase in the reduction in hyaluronic acid expression along with an increase in

incubation time. This shows that hyaluronic acid undergoes upregulation and then degrades due to exposure to high doses of glucose. At initial incubation, there is dominance in the synthesis of hyaluronic acid through hyaluronan synthase, uridine-diphosphate (UDP)-glucose precursor, and the influence of Mg catalyst (Viola et al., 2008; Gallo et al., 2019). In the final incubation, there is dominance in the direction of degradation by hyaluronidase (Dhulekar and Simionescu, 2018). Previous studies have shown that hvaluronidase can be activated by hyperglycemia, thereby contributing to decreased glycocalyx volume (Dogné et al., 2016). In hyperglycemia, thrombospondin-2 enables HAS-2, thereby increasing plasma levels of hyaluronic acid in diabetic patients (Vigetti et al., 2014). Reactive oxygen compounds are also involved in this process (Eshaq, Wright and Harris, 2014). Reactive oxygen compounds not only have a direct effect on hyaluronic acid but can trigger activation of proteolysis through metalloproteinase matrix and inactivation of endogenous protease inhibitors. This study extends the previous model that hyaluronic acid dearadation occurs in diabetic mice (Wolff and Dean, 1987).

For heparan sulfate, significant degradation was found at 48 hours incubation. Heparan sulfate degradation is played by sulfatase and hepaticase (Patel, Pineda and Hoffman, 2017) as well as pH and mechanical stimulus (Reitsma et al., 2007). For high-dose glucose exposure, there was a significant increase in the expression of syndecan from 48 hours of incubation. This shows the upregulation of syndecan in high glucose levels. We hypothesize that this increased expression is a mechanism of endothelial cell protection against glycocalyx degradation. The glycocalyx function depends on distribution and stability (Zeng et al., 2014). This study is not consistent with previous findings that exposure to high glucose levels can reduce syndecan (Han and Hiebert, 2013).

Effect of glucose dosage on glycocalyx

Under normal conditions, hyaluronic acid is in a massive molecular form (> 500 kDa) and is antiinflammatory. In inflammatory and disease conditions, hyaluronic acid is degraded into small fragments molecular weight and proinflammatory (Hull et al., 2015). At 12 and 72 hours of incubation, the higher the dose, the lower the expression of hyaluronic acid in HUVECs cells. This indicates that the 12 and 72 hours incubation, the higher the glucose dose, the greater the degradation of hyaluronic acid. Hyaluronic acid degradation results from gradual local degradation (Saegusa, Isaji and Kawarada,

2002; Leskova et al., 2019), or due to enzymatic activity, including hyaluronidase (Chajara et al., 2000; Chowdhury et al., 2016), thrombin, cathepsin B, proteinase-3, and plasmin (Becker et al., 2015) or due to oxidative stress (Jiang, Liang and Noble, 2011). Hyaluronidase breaks down sizeable molecular weight hyaluronic acid into small molecular weight, but its enzyme activity does not occur in little molecular weight hyaluronic acid (Rügheimer et al., 2009). This study extends previous findings that the degradation of hyaluronic acid in diabetic mice (Leskova et al., 2019), through the activation of hyaluronidase (Dogné et al., 2016), as well as the role of reactive oxygen compounds (Eshaq, Wright and Harris, 2014).

Glycocalyx degradation does not occur in heparan sulfate. Interestingly, syndecan was found to be significantly increased. This shows the upregulation of syndecan in high glucose levels. We hypothesize that this increased expression is a mechanism of endothelial cell protection against glycocalyx degradation. The glycocalyx function depends on distribution and stability (Zeng et al., 2014). This study is not following previous findings that high glucose levels can reduce syndecan expression in endothelial cells (Han and Hiebert, 2013).

Comparison of single and intermittent doses

Previous studies have suggested that fluctuations in glucose concentrations (peak hyperglycemia and hypoglycemia) stimulate the production of reactive oxygen compounds and cellular apoptosis more severe than constant high sugar levels (Risso et al., 2001; Quagliaro et al., 2003; Piconi et al., 2006). If endothelial cells are exposed to stable hyperglycemia, the production of reactive oxygen compounds produced will also be permanent. This will trigger initiation, propagation, and termination reactions. At the time of the termination reaction, every radical produced will always form a neutral compound when the formation of other radicals is stable. When the production of reactive oxygen compounds due to hyperglycemia is fluctuating, the termination reaction is incomplete, and radical compounds are formed and trigger oxidative stress.

In this study, the expression of hyaluronic acid decreased significantly with glucose exposure at dose 22 compared with a dose of glucose 0 mM or intermittent dose compared with glucose at a dose of 0 mM. This reduction is more significant in high-dose exposure than in intermittent doses. This indicates three things: First, high-dose glucose exposure triggers more severe degradation of hyaluronic acid than intermittent

glucose exposure. Hyaluronic acid degradation through gradual local degradation (Saegusa, Isaji and Kawarada, 2002; Leskova et al., 2019), hyaluronidase activity (Chajara et al., 2000; Chowdhury et al., 2016; Marini et al., 2018), thrombin, cathepsin B, proteinase-3, and plasmin (Becker et al., 2015), or oxidative stress (Jiang, Liang and Noble, 2011). This finding is not consistent with previous studies that functional injury to HUVECs cells is more severe due to variability than hyperglycemia alucose (Abdelzaher et al., 2016; Guo et al., 2016). Second, the low expression of hyaluronic acid also indicates the ability of endothelial cells to survive oxidative stress. Previous research stated that high molecular hyaluronic acid as an antioxidant (Ye et al., 2012). Scavenging and antioxidant activity are higher in low molecular compared to high molecular hyaluronic acid (Ke et al., 2011). On the other hand, the release of hyaluronic acid from endothelial cells is thought to contribute to the development of complications of diabetes mellitus (Infante et al., 2017).

For heparan sulfate, there is an increase in expression at doses of 22 mM and intermittent doses compared to physiological doses. A more significant improvement was found for intermittent doses. This indicates that exposure to high doses of glucose and intermittent doses triggers the upregulation of heparan sulfate. We suspect that this increase in heparan sulfate is a compensatory mechanism due to the degradation of hyaluronic acid.

For syndecan, there was a significant increase in dose 22 mM compared to doses 5 mM and 0 mM. For intermittent doses, there was no difference compared to treatments 5 and 0. This indicates the upregulation of syndecan in high-dose glucose exposure(Guo et al., 2016). We hypothesize that this increased expression is a mechanism of endothelial cell protection against glycocalyx degradation. The glycocalyx function depends on distribution and stability (Zeng et al., 2014). This study is not by previous findings that high glucose levels can reduce syndecan expression in endothelial cells (Han and Hiebert, 2013; Zhu et al., 2019).

Conclusion

In conclusions, in conditions without glucose exposure and physiological glucose exposure, glycocalyx has a dynamic expression. High glucose exposure eliminates the dynamics of hyaluronic acid and syndecan expression. Single-dose exposure triggers a decrease in hyaluronic acid expression compared to intermittent exposure, and this is the opposite of heparan sulfate exposure.

Declarations Funding statement

This research was not funded by any institution, but was funded by the individual himself.

Competing interest statement

The authors declare no conflict of interest.

Additional information

No additional information is available for this paper.

References

- Abdelzaher, L. A. et al. 2016. Astaxanthin alleviates oxidative stress insults-related derangements in human vascular endothelial cells exposed to glucose fluctuations. Life Sciences. 150, 24–31. doi: 10.1016/i.lfs.2016.02.087.
- Bammert, T. D. et al. 2017. High glucose derived endothelial microparticles increase active caspase-3 and reduce microRNA-Let-7a expression in endothelial cells. Biochem and Biophysic Res Comm. 493(2), pp. 1026–1029. doi: 10.1016/j.bbrc.2017.09.098.
- Barakat, A. I. 2008. Dragging along: The glycocalyx and vascular endothelial cell mechanotransduction. Circulation Res. 102(7), 747–748. doi: 10.1161/CIRCRESAHA.108.174839.
- Becker, B. F. et al. 2015. Degradation of the endothelial glycocalyx in clinical settings: Searching for the sheddases. British. J. Clin. Pharmacol. 80(3), 389–402. doi: 10.1111/bcp.12629.
- Broekhuizen, L. N. et al. 2010. Effect of sulodexide on endothelial glycocalyx and vascular permeability in patients with type 2 diabetes mellitus. Diabetologia. 53(12), 2646–2655. doi: 10.1007/s00125-010-1910-x.
- Chajara, A. et al. 2000. Increased hyaluronan and hyaluronidase production and hyaluronan degradation in injured aorta of insulin-resistant rats. Arteriosclerosis. Throm. Vascul. Biolog. 20(6), 1480–1487. doi: 10.1161/01.ATV.20.6.1480.
- Chowdhury, B. et al. 2016. Hyaluronidase 2 (HYAL2) is expressed in endothelial cells, as well as some specialized epithelial cells, and is required for normal hyaluronan catabolism. Histochemistry and Cell Biology. 145(1), 53–66. doi: 10.1007/s00418-015-1373-8.
- Dalan, R. et al. 2014. Vitamin D and the endothelium: Basic, translational and clinical research updates. IJC Metabol and Endocrine. 4, 4–17. doi: 10.1016/j.ijcme.2014.06.003.
- DeAngelis, P. L. 1996. Enzymological characterization of the Pasteurella multocida hyaluronic acid synthase. Biochem. 35(30), 9768–

- 9771. doi: 10.1021/bi960154k.
- Deguchi, Y. and Miyazaki, K. 2010. Antihyperglycemic and anti-hyperlipidemic effects of guava leaf extract. Nutrition and Metabolism. 7, pp. 1–10. doi: 10.1186/1743-7075-7-9.
- Dhulekar, J. and Simionescu, A. 2018. Challenges in vascular tissue engineering for diabetic patients. Acta Biomaterialia. 70, 25–34. doi: 10.1016/j.actbio.2018.01.008.
- Dogné, S. et al. 2016. Hyaluronidase I deficiency preserves endothelial function and glycocalyx integrity in early streptozotocin-induced diabetes. Diabetes. 65(9), 2742–2753. doi: 10.2337/db15-1662.
- Eshaq, R. S., Wright, W. S. and Harris, N. R. 2014. Oxygen delivery, consumption, and conversion to reactive oxygen species in experimental models of diabetic retinopathy. Redox Biology. 2(1), 661–666. doi: 10.1016/i.redox.2014.04.006.
- Fels, J. et al. 2014. Nanomechanics of vascular endothelium. Cell and Tissue Research. 355(3), 727–737. doi: 10.1007/s00441-014-1853-5.
- Ferrè, S. et al. 2010. Magnesium deficiency promotes a pro-atherogenic phenotype in cultured human endothelial cells via activation of NFkB. Biochimica et Biophysica Acta - Molecular Basis of Disease, 1802(11), 952–958. doi: 10.1016/j.bbadis.2010.06.016.
- Fitzgerald, M. L. et al. 2000. Shedding of syndecan-I and -4 ectodomains is regulated by multiple signaling pathways and mediated by a TIMP-3-sensitive metalloproteinase. J of Cell Biology. 148(4), 811–824. doi: 10.1083/icb.148.4.811.
- 17. Fox, C. S. et al. 2015. Update on prevention of cardiovascular disease in adults with type 2 diabetes mellitus in light of recent evidence: A scientific statement from the American Heart Association and the American diabetes association. Diabetes Care. 38(9), 1777–1803. doi: 10.2337/dci15-0012.
- Fu, B. M. and Tarbell, J. M. 2013. Mechanosensing and transduction by endothelial surface glycocalyx: Composition, structure, and function. Wiley Interdisciplinary Reviews: Systems Biology and Medicine. 5(3), 381–390. doi: 10.1002/wsbm.1211.
- Gallo, N. et al. 2019. Hyaluronic acid for advanced therapies: Promises and challenges. European Polymer Journal. 117(May), 134–147. doi: 10.1016/j.eurpolymj.2019.05.007.
- Georgescu, A. et al. 2011. The promise of EPC-based therapies on vascular dysfunction in diabetes. European Journal of Pharmacology. 669(1–3), 1–6. doi: 10.1016/j.ejphar.2011.07.035.
- Guo, J. et al. 2016. MiR-1273g-3p participates in acute glucose fluctuation-induced autophagy, dysfunction, and proliferation attenuation in human umbilical vein endothelial cells. American

- Journal of Physiology Endocrinology and Metabolism. 310(9), E734–E743. doi: 10.1152/ajpendo.00444.2015.
- Han, J. and Hiebert, L. M. 2013. Alteration of endothelial proteoglycan and heparanase gene expression by high glucose, insulin and heparin. Vascular Pharmacology. 59(3–4), 112–118. doi: 10.1016/j.vph.2013.08.001.
- Home, P. 2005. Contributions of basal and postprandial hyperglycaemia to micro- and macrovascular complications in people with type 2 diabetes. Current Medical Research and Opinion. 21(7), 989–998. doi: 10.1185/030079905X49662.
- 24. Hossain, P., Kawar, B. and El Nahas, M. 2007. Obesity and diabetes in the developing world - A growing challenge. New England Journal of Medicine. 356(3), 213–215. doi: 10.1056/NEJMp068177.
- Huang, G., Chen, J. and Chen, J. 2018. Accepted Manuscript'.
- Hull, R. L. et al. 2015. Hyaluronan: A Mediator of Islet Dysfunction and Destruction in Diabetes?. J of Histochemistry and Cytochem. 63(8), 592– 603. doi: 10.1369/0022155415576542.
- Infante, T. et al. 2017. In vivo and In vitro analysis in coronary artery disease related to type 2 diabetes. Frontiers in Endocrinology. 8(AUG). doi: 10.3389/fendo.2017.00209.
- Jiang, D., Liang, J. and Noble, P. W. 2011. Hyaluronan as an immune regulator in human diseases. Physiological Reviews. 91(1), 221–264. doi: 10.1152/physrev.00052.2009.
- Jing, Z. et al. 2016. Downregulation of Syndecan-I induce glomerular endothelial cell dysfunction through modulating internalization of VEGFR-2', Cellular Signalling. 28(8), pp. 826–837. doi: 10.1016/j.cellsig.2016.04.001.
- Ke, C. et al. 2011. Antioxidant acitivity of low molecular weight hyaluronic acid. Food and Chemical Toxicology. 49(10), pp. 2670–2675. doi: 10.1016/j.fct.2011.07.020.
- 31. Khanmohammadi, M., Sakai, S. and Taya, M. 2017. Impact of immobilizing of low molecular weight hyaluronic acid within gelatin-based hydrogel through enzymatic reaction on behavior endothelial cells. enclosed of International of Journal **Biological** Macromolecules. 308-316. 97. doi: 10.1016/j.ijbiomac.2016.12.088.
- Kristensen, S. L. et al. 2016. Risk Related to Pre-Diabetes Mellitus and Diabetes Mellitus in Heart Failure with Reduced Ejection Fraction: Insights from Prospective Comparison of ARNI with ACEI to Determine Impact on Global Mortality and Morbidity in Heart Failure Trial. Circulation: Heart Failure. 9(1), 1–12. doi: 10.1161/CIRCHEARTFAILURE.115.002560.
- 33. Leskova, W. et al. 2019. Effect of diabetes and hyaluronidase on the retinal endothelial

- glycocalyx in mice. Experimental Eye Research. 179, 125–131. doi: 10.1016/j.exer.2018.11.012.
- Li, R. et al. 2016. Syndecan-4 shedding impairs macrovascular angiogenesis in diabetes mellitus. Biochemical and Biophysical Research Communications. 474(1), pp. 15–21. doi: 10.1016/j.bbrc.2016.03.112.
- Marini, G. et al. 2018. The influence of hyperglycemia on the remodeling of urethral connective tissue in pregnant rats. European Journal of Obstetrics and Gynecology and Reproductive Biology. 221, 81–88. doi: 10.1016/j.ejogrb.2017.12.032.
- Nieuwdorp, M. et al. 2006. and Coagulation Activation In Vivo. Diabetes. 55(February), 480– 486
- Patel, V. N., Pineda, D. L. and Hoffman, M. P. 2017. The function of heparan sulfate during branching morphogenesis. Matrix Biology. 57–58, 311–323. doi: 10.1016/j.matbio.2016.09.004.
- Piconi, L. et al. 2006. Constant and intermittent high glucose enhances endothelial cell apoptosis through mitochondrial superoxide overproduction. Diabetes/Metabolism Research and Reviews. 22(3), pp. 198–203. doi: 10.1002/dmrr.613.
- Quagliaro, L. et al. 2003. Intermittent High Glucose Enhances Apoptosis Related to Oxidative Stress in Human Umbilical Vein Endothelial Cells: The Role of Protein Kinase C and NAD(P)H-Oxidase Activation. Diabetes. 52(11), 2795–2804. doi: 10.2337/diabetes.52.11.2795.
- Quincozes-Santos, A. et al. 2017. Fluctuations in glucose levels induce glial toxicity with glutamatergic, oxidative and inflammatory implications. Biochimica et Biophysica Acta -Molecular Basis of Disease. 1863(1), 1–14. doi: 10.1016/j.bbadis.2016.09.013.
- 41. Reitsma, S. et al. 2007. The endothelial glycocalyx: Composition, functions, and visualization', Pflugers Archiv European Journal of Physiology, 454(3), pp. 345–359. doi: 10.1007/s00424-007-0212-8.
- Ricks, J. et al. 2012. Glycemic control and cardiovascular mortality in hemodialysis patients with diabetes: A 6-year cohort study. Diabetes. 61(3), pp. 708–715. doi: 10.2337/db11-1015.
- Risso, A. et al. 2001. Intermittent high glucose enhances apoptosis in human umbilical vein endothelial cells in culture. American Journal of Physiology - Endocrinology and Metabolism. 281(5 44-5), 924–930. doi: 10.1152/ajpendo.2001.281.5.e924.
- Rügheimer, L. et al. 2009. Hyaluronan synthases and hyaluronidases in the kidney during changes in hydration status. Matrix Biology. 28(7), 390– 395. doi: 10.1016/j.matbio.2009.07.002.
- 45. Saegusa, S., Isaji, S. and Kawarada, Y. 2002. Changes in serum hyaluronic acid levels and

- expression of CD44 and CD44 mRNA in hepatic sinusoidal endothelial cells after major hepatectomy in cirrhotic rats. World Journal of Surgery. 26(6), 694–699. doi: 10.1007/s00268-001-0292-0.
- Saito, A. 2015. Heparin cofactor II is degraded by heparan sulfate and dextran sulfate. Biochemical and Biophysical Research Communications. 457(4), 585–588. doi: 10.1016/j.bbrc.2015.01.028.
- Savery, M. D. et al. 2013. The endothelial glycocalyx in syndecan-1 deficient mice. Microvascular Research. 87, 83–91. doi: 10.1016/j.mvr.2013.02.001.
- Sieve, I., Münster-Kühnel, A. K. and Hilfiker-Kleiner, D. 2018. Regulation and function of endothelial glycocalyx layer in vascular diseases. Vascular Pharmacology. 100, 26–33. doi: 10.1016/j.vph.2017.09.002.
- Subramanian, S. V., Fitzgerald, M. L. and Bernfield, M. 1997. Regulated shedding of syndecan-1 and -4 ectodomains by thrombin and growth factor receptor activation. Journal of Biological Chemistry. 272(23), 14713–14720. doi: 10.1074/jbc.272.23.14713.
- Tarbell, J. M. and Pahakis, M. Y. 2006. Mechanotransduction and the glycocalyx. Journal of Internal Medicine. 259(4), 339–350. doi: 10.1111/j.1365-2796.2006.01620.x.
- Vigetti, D. et al. 2014. Epigenetics in extracellular matrix remodeling and hyaluronan metabolism. FEBS Journal. 281(22), 4980–4992. doi: 10.1111/febs.12938.
- Viola, M. et al. 2008. Molecular control of the hyaluronan biosynthesis. Connective Tissue Research. 49(3–4), III-II4. doi: 10.1080/03008200802148405.
- 53. Wang, J. bo et al. 2009. Insulin increases shedding of syndecan-1 in the serum of patients with type 2 diabetes mellitus', Diabetes Research and Clinical Practice, 86(2), pp. 83–88. doi: 10.1016/j.diabres.2009.08.002.
- Wolff, P. and Dean, T. 1987. A section of, 232, pp. 283–293.
- Yang, H. et al. 2019. Berberine inhibits low shear stress-induced glycocalyx degradation via modulating AMPK and p47phox/Hyal2 signal pathway. European Journal of Pharmacology. 856(January). doi: 10.1016/j.ejphar.2019.172413.
- Ye, J. et al. 2012. High molecular weight hyaluronan decreases oxidative DNA damage induced by EDTA in human corneal epithelial cells. Eye (Basingstoke). 26(7), 1012–1020. doi: 10.1038/eye.2012.89.
- 57. Zeng, Y. et al. 2014. Sphingosine-I-phosphate protects endothelial glycocalyx by inhibiting syndecan-I shedding. American Journal of Physiology - Heart and Circulatory Physiology. 306(3). doi: 10.1152/ajpheart.00687.2013.
- 58. Zhu, T. et al. 2019. Ginsenoside RgI attenuates

high glucose-induced endothelial barrier dysfunction in human umbilical vein endothelial cells by protecting the endothelial glycocalyx. Experimental and Therapeutic Medicine. 3727–3733. doi: 10.3892/etm.2019.7378.

ORIGIN	IALITY REPORT				
SIMIL	3% ARITY INDEX	8% INTERNET SOURCES	5% PUBLICATIONS	3% STUDENT PA	PERS
PRIMAI	RY SOURCES				
1	www.karyailmiah.trisakti.ac.id Internet Source				5
2	Submitted to Fakultas Kedokteran Gigi Universitas Trisakti Student Paper				3
3	Rajkumar, K "Relationship between concentration of prostaglandins E and F in the regulation of ovum transport in rabbits", Prostaglandines and Medicine, 197906 Publication				29
4	Yuche Wu, Yanming Wang, Xinhua Nabi. "Protective effect of Ziziphora clinopodioides flavonoids against H2O2-induced oxidative stress in HUVEC cells", Biomedicine & Pharmacotherapy, 2019 Publication				29
5	M. O. Semin, S. A. Apryatin, I. V. Gmoshinskii, D. B. Nikityuk. "Comparative Analysis of JNK1 Expression in Liver Cells in Rats of Different Lines Receiving Excess of Easily Digested				2

Carbohydrates: Confocal Microscopy", Bulletin of Experimental Biology and Medicine, 2019

Publication

Exclude quotes On Exclude matches < 2%

Exclude bibliography On